



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<p>(51) International Patent Classification ⁶ : A61K 45/06, 31/37, 31/225, 31/195 // (A61K 31/37, 31:37, 31:225, 31:195) (A61K 31/225, 31:165) (A61K 31/195, 31:165)</p>	<p>A1</p>	<p>(11) International Publication Number: WO 96/22791</p> <p>(43) International Publication Date: 1 August 1996 (01.08.96)</p>
<p>(21) International Application Number: PCT/US96/00420</p> <p>(22) International Filing Date: 16 January 1996 (16.01.96)</p> <p>(30) Priority Data: 377,584 23 January 1995 (23.01.95) US</p> <p>(71) Applicant: OCTAMER, INC. [US/US]; Suite 255-A, 100 Shoreline Highway, Mill Valley, CA 94941 (US).</p> <p>(72) Inventors: KUN, Ernest; 8 Helens Lane, Mill Valley, CA 94941 (US). MENDELEYEV, Jerome; 40 Harbor Oak Drive, Tiburon, CA 94920 (US). KIRSTEN, Eva; 397 Imperial Way #302, Daly City, CA 94015 (US).</p> <p>(74) Agents: HALLUIN, Albert, P. et al.; Pennie & Edmonds, 1155 Avenue of the Americas, New York, NY 10036 (US).</p>		<p>(81) Designated States: AL, AM, AU, AZ, BB, BG, BR, BY, CA, CN, CZ, EE, FI, GE, HU, IS, JP, KG, KP, KR, KZ, LK, LR, LS, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, TJ, TM, TR, TT, UA, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AZ, BY, KG, KZ, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>
<p>(54) Title: NOVEL SYNERGISTIC COMPOSITIONS USEFUL AS ANTI-TUMOR OR ANTI-RETROVIRAL AGENTS</p> <p>(57) Abstract</p> <p>The subject invention provides for novel synergistic compositions useful for inactivating viruses or inducing apoptosis in tumor cells and for treating cancer or retroviral infections. Generally, the compositions comprise one or a plurality of ligands that oxidatively attack a zinc finger of pADPRT in combination with one or a plurality of agents selected from the group consisting of: agents that decrease cellular levels of glutathione and ligands that non-covalently bind to the nicotinamide site of pADPRT but do not effect zinc ejection from a zinc finger of pADPRT.</p>		

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NOVEL SYNERGISTIC COMPOSITIONS USEFUL AS ANTI-TUMOR OR ANTI-
RETROVIRAL AGENTS

CROSS REFERENCE

5 This is a continuation-in-part of co-pending U.S. Application Serial No. 08/076,313 filed June 11, 1993, which is a continuation-in-part of co-pending U.S. Application Serial No. 08/060,409 filed May 12, 1993; and co-pending U.S. Application Serial No. 08/087,566 filed July 2, 1993, which
10 is a continuation-in-part of co-pending U.S. Application 07/965,541 filed November 2, 1992, which is a continuation-in-part of co-pending U.S. Serial No. 07/893,429 filed June 4, 1992, which is a continuation-in-part of co-pending U.S. Serial No. 07/780,809 filed October 22, 1991.

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FIELD OF THE INVENTION

 The present invention relates generally to the fields of retroviral and cancer therapeutic and inactivating agents and to their use in treating retroviral infections and cancer.
20 More specifically, the present invention relates to novel synergistic compositions useful as potent, selective and safe anti-tumor or anti-retroviral agents. The synergistic compositions comprise one or a plurality of ligands that oxidatively attack a zinc finger of poly (ADP-ribose)
25 polymerase ("pADPRT") in combination with one or a plurality of agents selected from the group consisting of: agents that decrease cellular levels of glutathione ("GSH decreasing agents") and ligands that bind non-covalently to the nicotinamide site of pADPRT but do not eject zinc from a zinc
30 finger of pADPRT ("pADPRT-inhibitory ligands").

 The invention also relates generally to methods for treating tumorigenic and retroviral diseases using said synergistic compositions.

35

BACKGROUND OF THE INVENTION

Cancer and viral infections are a serious threat to modern society. Malignant cancerous growths, due to their unique characteristics, pose serious challenges for modern
5 medicine. These characteristics include: uncontrollable cell proliferation resulting in unregulated growth of malignant tissue, an ability to invade local and even remote tissues, lack of differentiation, lack of detectable symptoms and most significantly, the lack of effective therapy and prevention.

10 Cancer can develop in any tissue of any organ at any age. The etiology of cancer is not clearly defined but mechanisms such as genetic susceptibility, chromosome breakage disorders, viruses, environmental factors and immunologic disorders have all been linked to a malignant
15 cell growth and transformation.

Antineoplastic chemotherapy currently encompasses several groups of drugs including alkylating agents, purine antagonists and antitumor antibiotics. Alkylating agents alkylate cell proteins and nucleic acids preventing cell
20 replication, disrupting cellular metabolism and eventually leading to cell death. Typical alkylating agents are nitrogen mustard, cyclophosphamide and chlorambucil. Toxicities associated with alkylating agents treatment include nausea, vomiting, alopecia, hemorrhagic cystitis,
25 pulmonary fibrosis and an increased risk of development of acute leukemia.

Purine, pyrimidine and folate antagonists are cell cycle and phase specific and, in order to promote an anti-tumor effect, they require cells to be in the cell replication
30 cycle and in the DNA synthesis phase of replication. The purine antagonists such as 6-mercaptopurine or 6-thioguanidine inhibit de novo purine synthesis and interconversion of purines. The pyrimidine antagonists, such as cytarabine, 5-fluorouracil or floxuridine inhibit DNA
35 synthesis by inhibiting deoxycytidylate kinase and DNA polymerase.

Folate antagonists, e.g., methotrexates, bind tightly with the intracellular enzyme dihydrofolate reductase ultimately leading to cell death resulting from an inability to synthesize pyrimidines. Toxicities associated with the use of these compounds include alopecia, myelosuppression, vomiting, nausea, and cerebellar ataxia, among others.

Plant alkaloids such as vincristine, vinblastine or podophyllotoxins etoposide and teniposide generally inhibit mitosis and DNA synthesis and RNA dependent protein synthesis. Toxicities of these drugs are similar to those described above and include myopathy, myelosuppression, peripheral neuropathy, vomiting, nausea and alopecia.

Antitumor antibiotics such as doxorubicin, daunorubicin and actinomycin act as intercalators of DNA, preventing cell replication, inhibiting synthesis of DNA-dependent RNA and inhibiting DNA polymerase. Bleomycin causes scission of DNA and mitomycin acts as inhibitor of DNA synthesis by bifunctional alkylation. Toxicities of these antibiotics are numerous and severe and include necrosis, myelosuppression, anaphylactic reactions, anorexia, dose-dependent cardiotoxicity and pulmonary fibrosis.

Other compounds used for chemotherapeutical treatment of cancer are inorganic ions such as cisplatin, biologic response modifiers such as interferon, enzymes and hormones. All these compounds, similarly to those mentioned above, are accompanied by toxic adverse reactions.

Thus, it would be extremely advantageous to provide safe and non-toxic chemotherapeutic compositions which would effectively inhibit cancer cell proliferation and suppress neoplastic growth. The Merck Manual, 1218-1225 (1987), 15th Ed. Novel synergistic compositions of this invention provide such treatment.

Similar to cancer, the high degree of infectiousness and fast reproduction cycle of viruses within host organisms make viruses a nuisance and a health hazard.

There is no simple treatment of viral diseases. Viruses are not susceptible to antibiotics. The only available

treatment of viral diseases is chemotherapy utilizing viral replication inhibitors in host cells. The Merck Manual, 170, 14th Ed. (1982). Examples of these chemical agents are idoxuridine, acyclovir, ribavirin, vidarabine, gancyclovir, 5 adenine arabinoside (ABA-A) and AZT. These, and other viral replication inhibitors, however, are cytotoxic, hepatotoxic, neurotoxic, nephrotoxic and teratogenic. Virus Diseases, 1-6, Crown Publishers, N.Y. (1978).

Human immunodeficiency virus (HIV) infections known as 10 acquired immunodeficiency syndrome (AIDS), presently constitute a worldwide health hazard. HIV infections are almost always fatal due to a weakened immunoresistance, leading to opportunistic infections, malignancies and neurologic lesions.

15 There is no effective treatment for AIDS other than the treatment of the opportunistic infections, neoplasms and other complications. Available cytostatic (AZT) and antiviral (acyclovir) drugs are extremely toxic and cause severe adverse reactions. In addition, opioid drug addiction 20 coincides with the emergence of AZT resistance, Chuang et al., NIDA Res. Monograph: Problems of Drug Dependence 14:419 (1993), thereby reducing the efficacy of AZT treatment for opioid-addicted AIDS patients.

Thus it would be highly desirable to have available an 25 effective and yet nontoxic treatment of viral diseases, in particular, AIDS, and further for AZT-resistant patients or viral strains.

Cytomegalovirus (CMV), a dangerous co-infection of HIV, is a subgroup of highly infectious viruses having the 30 propensity for remaining latent in man. CMVs are very common among the adult population and as many as 90% of adults have been exposed to and experienced CMV infections. CMVs are normally present in body liquids such as blood, lymph, saliva, urine, feces, milk, etc. CMV infections may cause 35 abortion, stillbirth, postnatal death from hemorrhage, anemia, severe hepatic or CNS damage. Particularly dangerous are CMV infections afflicting AIDS patients, where CMV may

cause pulmonary, gastrointestinal or renal complications. There is no specific therapy for CMVs. CMV is resistant to acyclovir, and to other known antiviral drugs.

Thus, it would be extremely advantageous to have
5 available a drug which would effectively inhibit CMV infections.

Recently, a series of highly effective anti-tumor and anti-viral drugs were identified. These drugs include: substituted and unsubstituted 6-amino-1,2-benzopyrones which
10 are the subject of copending U.S. patent application Serial No. 08/237,969 filed on May 3, 1994, entitled "6-Amino-1,2-Benzopyrones Useful for Treatment of Viral Diseases;" 5-iodo-6-amino-1,2-benzopyrones and 5-iodo-6-nitroso-1,2-benzopyrones which are the subject of copending U.S. patent
15 applications Serial No. 07/600,593 filed on October 19, 1990 entitled "Novel 5-Iodo-6-Amino-1,2-Benzopyrones and Their Metabolites Useful as Cytostatic and Antiviral Agents" and Serial No. 08/021,989 filed on February 24, 1993 entitled "Novel 5-Iodo-6-Amino-1,2-Benzopyrones and Their Metabolites
20 Useful as Cytostatic Agents;" 3-nitrosobenzamides, 6-nitroso-1,2-benzopyrones and nitroso-1-(2H)-isoquinolinones which are the subject of copending U.S. patent applications Serial No. 07/780,809, 07/893,429 and 07/965,541 filed October 22, 1991, June 4, 1992 and November 2, 1992, respectively, and entitled
25 "Adenosine Diphosphoribose Polymerase Binding Nitroso Aromatic Compounds Useful As Retroviral Inactivating Agents, Anti-retroviral Agents and Anti-tumor Agents;" various iodo-nitro compounds and iodo-nitroso compounds, which are the subject of copending U.S. patent application Serial No.
30 08/060,409 filed on May 12, 1993, entitled "Novel Aromatic Nitro Compounds and Their Metabolites Useful as Anti-Viral and Anti-Tumor Agents," and U.S. patent application Serial No. 08/076,313 filed June 11, 1993, entitled "Novel Aromatic Nitro and Nitroso Compounds and Their Metabolites Useful as
35 Anti-Viral and Anti-Tumor Agents," the disclosures of which are incorporated herein by reference.

These drugs are of remarkably low toxicity, yet highly effective inhibitors of tumors and viral replication in cell cultures. Their therapeutic spectrum appears to be particularly useful for suppression and inhibition of cancer growth and the treatment of viral infections.

The mechanisms of action of poly (ADP ribose) transferase polymerizing ("pADPRT") CCHC-oxidizing ligands such as aromatic C-nitroso compounds have been recently elucidated. C-nitroso ligands exert cellular action on two levels. Kun et al., Biochemie, in press (1994). One level comprises inactivation of pADPRT by zinc ejection. Recently published experiments have shown that aromatic C-nitroso ligands of pADPRT preferentially destabilize one of the two zinc fingers of the enzyme coincidentally with a loss of enzymatic activity but not DNA binding capacity of the protein. Buki et al., FEBS Lett. 290:181-185 (1991). Based on the similarity to results obtained by site-directed mutagenesis, Gradwohl et al., Proc. Natl. Sci. USA 87:2990-2992 (1990), it appears that the primary attack of C-nitroso ligands occurred at zinc finger F1, Buki et al., FEBS Lett. 290:181-185 (1991). 6-nitroso-1,2 benzopyrone ("NOBP") and 3-nitrosobenzamide ("NOBA"), two C-nitroso compounds that inactivate pADPRT at one zinc finger site completely suppressed the proliferation of leukemic and other malignant human cells and subsequently produced cell death. Tumorcidal concentrations of the drugs were relatively harmless to normal bone marrow progenitor cells and to superoxide formation by neutrophil granulocytes. The cellular mechanisms elicited by the C-nitroso compounds consists of apoptosis due to DNA degradation by the nuclear calcium/magnesium dependent endonuclease. Rice, et al. Proc. Natl. Sci. USA 89:7703-7707 (1992). This endonuclease is maintained in a latent form by poly(ADP-ribosyl)ation, but inactivation of pADPRT by C-nitroso drugs de-represses the DNA-degrading activity.

In contrast to the "classical" zinc finger structures identified in a large number of transcription factors where

the zinc ligands are CCCC or CCHH, Klug and Rhodes, Trends in Biochem. Sci. 12:464-469 (1987), pADPRT contains two zinc chelates that are asymmetrical, i.e. the ligands are CCHC, Gradwohl et al., Proc. Natl. Acad. Sci. 87:2990-2994 (1990).

5 This seemingly minor modification of the chelate has significant chemical consequence with respect to its stability as it exists in the zinc finger within the protein molecule. Buki et al., FEBS Lett. 290:181-185 (1991). Whereas CCCC or CCHH zinc fingers require basic pH and

10 organomercurials to release zinc ion, Giedroc et al., J. Inorg. Biochem. 28:155-169 (1986), zinc ion exchange between the CCHC zinc fingers of pADPRT and external zinc under physiological circumstances, and oxidative destruction of the CCHC chelates by relatively mild oxidizing agents that are

15 made selective by also being ligands of pADPRT has been demonstrated, Buki et al., FEBS Lett. 290:181-185 (1991). This represents a significant biochemical correlation. The F1 finger, Gradwohl et al., Proc. Natl. Acad. Sci. 87:2990-2994 (1990), of pADPRT is first inactivated by the oxidative

20 ligands, resulting in a loss of DNA-stimulated pADPRT activity without major alterations of the DNA binding ability of pADPRT, but excess oxidant destroys both zinc fingers, Buki et al., FEBS Lett. 290:181-185 (1991).

The second level of activity comprises induction of an

25 pADPRT-degrading aminopeptidase that completely digests pADPRT, whereby the pADPRT-binding sites on DNA become available to endonucleolytic degradation. Kun et al., Biochemie, in press (1994). Auto-poly-ADP-ribosylation protects pADPRT from digestion of aminopeptidase, hence

30 inactivation of poly-ADP-ribosylation (as by aromatic C-nitroso ligands) is a prerequisite to proteolytic pADPRT degradation, leading to apoptosis. Therefore, pADPRT CCHC-oxidizing ligand, such as C-nitroso compounds, are effective anti-tumor compounds.

35 Retroviral nucleocapsid ("NC") proteins and their respective gag precursors from all strains of known retroviruses contain at least one copy of a zinc-binding

polypeptide sequence of the type Cys-X₂-Cys-X₄-His-X₄-Cys ("CCHC"), i.e., a zinc finger domain. Henderson et al., Biol. Chem. 256:8400-8406 (1981). This CCHC sequence is essential for maintaining viral infectivity, Gorelick et al., 5 Proc. Natl. Acad. Sci. USA 85:8420-8424 (1988) and Gorelick et al., J. Virol. 64:3207-3211 (1990), therefore, it represents an attractive target for viral chemotherapy. The HIV-1 gag proteins function by specifically binding to the HIV-1 RNA, anchoring it to the cell membrane for budding of 10 viral particles. Meric et al., J. Virol. 63:1558-1658 (1989); Gorelick et al., Proc. Natl. Acad. Sci. USA 85:8420-8424 (1988); Aldovini et al., J. Virol. 64:1920-1926 (1990); and Lever et al., J. Virol. 63:4085-4087 (1989). Site-directed mutagenesis studies demonstrated that modification 15 of Cys or His residues results in defective viral RNA packaging and noninfectious viral particles are formed. Aldovini et al., J. Virol. 64:1920-1926 (1990) and Lever et al., J. Virol. 63:4085-4087 (1989).

Based on the occurrence of CCHC zinc binding sites in 20 both retroviral nucleocapsid and gag-precursor proteins and in poly(ADP-ribose) polymerase it was reasoned that pADPRT CCHC-oxidizing ligands may also have anti-retroviral effects. Recently it was demonstrated that NOBA and NOBP inhibit infection of human immunodeficiency virus HIV-1 in human 25 lymphocytes and also eject zinc from isolated HIV-1 NC zinc fingers and from intact HIV-1 virions. U.S. Serial No. 0/087,566 filed July 2, 1993; U.S. Serial No. 08/076,313 filed June 11, 1993; Rice et al., Nature 361:473-475 (1993); Rice et al., Proc. Natl. Acad. Sci. 90:9721-9724 (1993); 30 Chuang et al., FEBS Lett. 326:140-144 (1993); and Wondrak et al., J. Biol. Chem. in press (1994). The zinc-ejected HIV-1 virions exhibit complete loss of infectivity in human lymphocytes. U.S. Serial No. 0/087,566 filed July 2, 1993 and U.S. Serial No. 08/076,313 filed June 11, 1993. It was 35 of special interest that the infectious cycle of AZT-resistant rhesus macaque (MMU-23740)-derived SIV was equally inhibited by NOBA, Chuang et al., FEBS Lett. 326:140-144

(1993), an observation that is particularly relevant to opioid drug abusers where addiction coincides with the emergence of AZT resistance, Chuang et al., NIDA Res. Monograph: Problems of Drug Dependence 14:419 (1993).

5 Therefore, the pADPRT CCHC-oxidizing ligands are effective anti-viral compounds.

While these C-nitroso pADPRT CCHC-oxidizing ligands have been found to be quite effective in in vitro tests, they are relatively water insoluble at physiological pH, exhibit
10 limited stability and limited predictability of delivery to the affected cells due to their solubility and stability characteristics. In addition, the relative chemical instability of aromatic C-nitroso pADPRT CCHC-oxidizing ligands towards glutathione ("GSH") is a distinct
15 disadvantage for their direct application in vivo. Recently precursor molecules which generate active C-nitroso compounds in vivo have been prepared to overcome some of these difficulties.

One recently prepared precursor, a cysteine sulfinic
20 adduct of 3-nitrosobenzamide, releases the C-nitroso molecule at weakly basic conditions. Kun et al., U.S. Patent No. 5,262,564 issued on November 16, 1993. This sulfinic adduct is an effective C-nitroso donor molecule in cell cultures.
Id.

25 As an alternative method, stable precursor C-nitro molecules which serve as pro-drugs for the active C-nitroso pADPRT CCHC-oxidizing ligands were recently prepared. U.S. patent application Serial No. 08/076,313 filed on June 11, 1993, which is incorporated herein by reference. A prototype
30 of these pro-drugs, 4-iodo-3-nitrobenzamide ("INO₂BA") has been shown to be stable, having a shelf life of at least one year. Mendeleyev et al., "Chemotherapeutic Activity of 4-Iodo-3-Nitrobenzamide I: Metabolic Reduction to the 3-Nitroso Derivative and Induction of Apoptosis in Tumor Cells
35 in Culture," submitted (1994) (hereinafter "Mendeleyev et al. (1994).")

The aromatic iodo-nitro pro-drugs are slowly but steadily enzymatically reduced within cells to the active aromatic iodo-nitroso compounds such that tumor cell apoptosis or inactivation of viral replication is induced.

5 Id. See also Chuang et al., Biochemical Pharmacology, submitted (1995). These aromatic iodo-nitro compounds thus provide a ready source of in vivo anti-tumor C-nitroso compounds.

On the benzene ring, ortho-iodo-substitution has been
10 shown to facilitate the electrochemical reduction of the nitro group. Fry, The Chemistry of Amino, Nitroso and Nitro Compounds and Their Derivatives. Part I., S. Patai, ed., 319-335, John Wiley and Sons, NY (1982). Recently the fluoro-, chloro- and bromo- analogues of INO₂BA were tested
15 for anti-tumor activity. These analogues were less effective than INO₂BA. Mendeleyev et al. (1994). Unsubstituted nitrobenzamides are at least 10-15 times less effective than INO₂BA. Id. Thus, it is highly probable that the metabolic reduction of aromatic iodo-nitro compounds in cells is
20 activated by the presence of the iodo group ortho to the nitro group. Id.

It is known that glutathione adds to aromatic nitroso compounds to form labile semimercaptal adducts. Eyer, Chem. Biol. Interactions 24:227-239 (1979); Umemoto et al., Chem. Biol. Interactions 68:57-69 (1988); Ellis et al., Chem. Biol. Interactions 82:151-163 (1992). The semimercaptals undergo oxygen atom migration to yield sulfinimides that hydrolyze to the corresponding aromatic amines plus glutathione sulfinic acid. In addition, it has recently been shown that both NOBA
25 and 4-iodo-nitrosobenzamide ("INOBA") are rapidly reduced to the corresponding hydroxyl amines by GSH and ascorbate in in vitro chemical tests. Since many cell types produce GSH, these reduction pathways can quickly reduce the active C-nitroso compounds available in vivo, thereby diminishing the
30 efficacy of the C-nitroso compounds in vivo.
35

It was thus reasoned that agents which decrease intracellular levels of GSH ("GSH decreasing agents") would

decrease the rate at which the active C-nitroso ligands are reduced to the corresponding amines in the cell. Recently, it has been shown that DL-buthionine sulfoximine ("BSO"), an agent that inhibits GSH biosynthesis, Meister, Pharmacology and Therapeutics 51:155-194 (1991), exhibits synergistic apoptosis-inducing potency when administered in combination with pADPRT CCHC-oxidizing ligands in several types of cancer cells. Mendeleyev et al. (1994). BSO also exhibited synergistic anti-viral activity when administered in combination with pADPRT CCHC-oxidizing ligands in cells infected with SIV. Chuang et al., "Chemotherapeutic Activity of 4-Iodo-3-Nitrobenzamide and 5-Iodo-6-Amino-Benzopyrone on Simian Immunodeficiency Virus (SIV) in Cultures of CEM x174 Cells," submitted (1994) (hereinafter "Chuang et al. (1994)").

It is therefore a primary object of this invention to provide stable, soluble, non-toxic, highly effective antineoplastic and/or anti-viral synergistic compositions that inactivate pADPRT and NC p7 zinc fingers oxidatively. pADPRT ligands that oxidize CCHC zinc fingers are not unique inhibitors of pADPRT. Inhibitory ligands of pADPRT that do not attack zinc have been identified. Such a prototype ligand, 6-amino-1,2-benzopyrone has been shown to have anti-tumor effects. Hakam et al., FEBS Lett. 193:1-4 (1991). These ligands bind non-covalently to the nicotinamide site and internal DNA-seeking domains of pADPRT. Bauer et al., manuscript in preparation (1994). In contrast, CCHC-oxidizing ligands also bind non-covalently to the nicotinamide site, but initially attack the F1 zinc finger oxidatively. Id. The respective molecular pharmacological action of both of these types of pADPRT-inhibitory ligands has been studied in a bovine endothelial cell line that has been transfected and transformed with E-ras and, by cellular cloning, the tumorigenic phenotype has been isolated. Id. Synergistic cytotoxic effects of pADPRT CCHC-oxidizing ligands in combination with non-covalent pADPRT-inhibitory

ligands has been demonstrated. U.S. Serial No. 08/076,313 filed June 11, 1993.

pADPRT ligands that oxidize CCHC zinc fingers are also not unique as potential anti-HIV drugs at the DNA level.

- 5 Ligands of pADPRT that do not attack zinc fingers were also shown to be anti-retroviral, Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991), by mechanisms that relate to the drug-induced binding of pADPRT to either the reverse transcriptase template, Buki et al., Biochem. Biophys. Res.
10 Commun. 180:496-503 (1991), or probably to transcription factors required for retroviral synthesis.

A prototypical molecule, 5-iodo-6-amino-1,2-benzopyrone ("IABP") blocked the infectious cycle of HIV as tested in various cell lines, Cole et al., Biochem. Biophys. Res.

- 15 Commun. 180:504-514 (1991), but had no influence on the CCHC zinc fingers of pADPRT or on the p7 NC protein of retroviruses. Thus, reversible inhibition of pADPRT as well as covalent inactivation of pADPRT by zinc ejection can abrogate HIV or SIV infection, in addition to a direct effect
20 of the pADPRT CCHC-oxidizing ligands on the retroviral zinc fingers of the virion p7, Rice et al., Nature 361:473-475 (1993) and Wondrak et al., J. Biol. Chem. 269:21948-21950 (1994), pointing to a hitherto unrecognized role of pADPRT in the life cycle of retroviruses.

- 25 Based on this newly recognized role of pADPRT on the retroviral life cycle and the observance of synergy between pADPRT CCHC-oxidizing ligands in combination with non-covalent pADPRT-inhibitory ligands in cancer cells, it was reasoned that non-covalent ligands of pADPRT in combination
30 with CCHC-oxidizing ligands would increase anti-retroviral activity as compared to such ligands acting independently.

- It is therefore also a primary objective of this invention to provide non-toxic, highly effective anti-tumor and/or anti-viral synergistic compositions that inactivate
35 pADPRT and NC p7 zinc fingers oxidatively, and that also inhibit pADPRT non-covalently.

SUMMARY OF THE INVENTION

The subject invention provides for novel synergistic anti-tumor and/or anti-retroviral compositions comprising one or a plurality of pADPRT CCHC-oxidizing ligands in
 5 combination with one or a plurality of agents selected from the group consisting of: GSH decreasing agents and non-covalent pADPRT-inhibitory ligands.

In one embodiment the subject invention provides for
 10 novel synergistic anti-tumor and/or anti-retroviral compositions comprising one or a plurality of pADPRT CCHC-oxidizing ligands such as nitro-estradiol, iodo-nitro-estradiol, nitroso-estradiol, iodo-nitroso-estradiol, nitro-estrone, iodo-nitro-estrone, nitroso-estrone, iodo-nitroso-
 15 estrone, nitro-estriol, iodo-nitro-estriol, nitroso-estriol, iodo-nitroso-estriol, nitro-equilenin, iodo-nitro-equilenin, nitroso-equilenin, iodo-nitroso-equilenin, nitro-equilin, iodo-nitro-equilin, nitroso-equilin, iodo-nitroso-equilin, nitro-diethylstilbestrol, iodo-nitro-diethylstilbestrol,
 20 nitroso-diethylstilbestrol, iodo-nitroso-diethylstilbestrol, nitrobenzamides, nitrosobenzamides, iodo-nitro-benzamides, iodo-nitrosobenzamides, 4-iodo-3-nitrobenzamide, 4-iodo-3-nitrosobenzamide, 2-iodo-5-nitrobenzamide, 2-iodo-5-nitrosobenzamide, nitro-1,2-benzopyrones, nitroso-1,2-
 25 benzopyrones, 6-nitro-1,2-benzopyrones, 6-nitroso-1,2-benzopyrones, iodo-nitro-1,2-benzopyrones, iodo-nitroso-1,2-benzopyrones, 4-iodo-3-nitro-1,2-benzopyrone, 4-iodo-3-nitroso-1,2-benzopyrone, 5-iodo-6-nitro-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, nitro-isoquinolinones, iodo-
 30 nitro-isoquinolinones, nitroso-isoquinolinones, iodo-nitroso-isoquinolinones, 5-nitro-1(2H)-isoquinolinone, iodo-5-nitro-1(2H)-isoquinolinones, 5-nitroso-1(2H)-isoquinolinone, iodo-5-nitroso-1(2H)-isoquinolinones, 7-nitro-1(2H)-isoquinolinone, iodo-7-nitro-1(2H)-isoquinolinone, 7-nitroso-
 35 1(2H)-isoquinolinone, iodo-7-nitroso-1(2H)-isoquinolinones, 8-nitro-1(2H)-isoquinolinone, iodo-8-nitro-1(2H)-isoquinolinone, 8-nitroso-1(2H)-isoquinolinone, iodo-8-

nitroso-1(2H)-isoquinolinones and their homologues, in combination with a one or a plurality of GSH decreasing agents such as BSO and diethylmaleate.

- 5 In another embodiment the subject invention also provides for novel synergistic anti-tumor and/or anti-viral compositions comprising one or a plurality of pADPRT CCHC-oxidizing ligands such as nitro-estradiol, iodo-nitro-estradiol, nitroso-estradiol, iodo-nitroso-estradiol, nitro-
 10 estrone, iodo-nitro-estrone, nitroso-estrone, iodo-nitroso-estrone, nitro-estriol, iodo-nitro-estriol, nitroso-estriol, iodo-nitroso-estriol, nitro-equilenin, iodo-nitro-equilenin, nitroso-equilenin, iodo-nitroso-equilenin, nitro-equilin, iodo-nitro-equilin, nitroso-equilin, iodo-nitroso-equilin,
 15 nitro-diethylstilbestrol, iodo-nitro-diethylstilbestrol, nitroso-diethylstilbestrol, iodo-nitroso-diethylstilbestrol, nitrobenzamides, nitrosobenzamides, iodo-nitro-benzamides, iodo-nitrosobenzamides, 4-iodo-3-nitrobenzamide, 4-iodo-3-nitrosobenzamide, 2-iodo-5-nitrobenzamide, 2-iodo-5-
 20 nitrosobenzamide, nitro-1,2-benzopyrones, nitroso-1,2-benzopyrones, 6-nitro-1,2-benzopyrones, 6-nitroso-1,2-benzopyrones, iodo-nitro-1,2-benzopyrones, iodo-nitroso-1,2-benzopyrones, 4-iodo-3-nitro-1,2-benzopyrone, 4-iodo-3-nitroso-1,2-benzopyrone, 5-iodo-6-nitro-1,2-benzopyrone, 5-
 25 iodo-6-nitroso-1,2-benzopyrone, nitro-isoquinolinones, iodo-nitro-isoquinolinones, nitroso-isoquinolinones, iodo-nitroso-isoquinolinones, 5-nitro-1(2H)-isoquinolinone, iodo-5-nitro-1(2H)-isoquinolinones, 5-nitroso-1(2H)-isoquinolinone, iodo-5-nitroso-1(2H)-isoquinolinones, 7-nitro-1(2H)-
 30 isoquinolinone, iodo-7-nitro-1(2H)-isoquinolinone, 7-nitroso-1(2H)-isoquinolinone, iodo-7-nitroso-1(2H)-isoquinolinones, 8-nitro-1(2H)-isoquinolinone, iodo-8-nitro-1(2H)-isoquinolinone, 8-nitroso-1(2H)-isoquinolinone, iodo-8-nitroso-1(2H)-isoquinolinones and their homologues, in
 35 combination with a one or a plurality of non-covalent pADPRT-inhibitory ligands such as 1,2-benzopyrones, benzamides, amino-1,2-benzopyrones, amino-benzamides, 5-amino-1,2-

benzopyrones, 3-aminobenzamides, amino-iodo-benzopyrones, amino-iodo-benzamides, 5-iodo-6-amino-1,2-benzopyrone, 4-iodo-3-aminobenzamides and their homologues.

- 5 The invention further provides for methods of treating cancer and viral diseases with such synergistic compositions.

Another aspect of the invention is to provide methods for inactivating viruses, especially retroviruses, in
10 biological material, e.g. blood, by adding one or a plurality of synergistic compositions to the biological material.

Another aspect of the invention is to provide methods for inactivating AZT-resistant viruses, in particular HIV and
15 SIV, by adding one or a plurality of synergistic compositions to a cell culture or mammalian host infected with an AZT-resistant virus.

Another aspect of the invention is to provide methods
20 for inducing apoptosis in a tumor cell by administering one or a plurality of synergistic compositions to a cell culture or mammalian host having one or a plurality of tumors.

In a preferred embodiment of the invention the pADPRT
25 CCHC-oxidizing ligands have an iodo group adjacent to the nitroso or nitro group. In an even more preferred mode the pADPRT CCHC-oxidizing ligands are aromatic C-nitro in vivo precursors of active C-nitroso ligands having an iodo group adjacent to a nitro group, including iodo-nitro-estradiol,
30 iodo-nitro-estrone, iodo-nitro-estriol, iodo-nitro-equilenin, iodo-nitro-equilin, iodo-nitro-diethylstilbestrol, nitrobenzamides, iodo-nitro-benzamides, 4-iodo-3-nitrobenzamide, 2-iodo-5-nitrobenzamide, nitro-1,2-benzopyrones, 6-nitro-1,2-benzopyrones, iodo-nitro-1,2-
35 benzopyrones, 4-iodo-3-nitro-1,2-benzopyrone, 5-iodo-6-nitro-1,2-benzopyrone, nitro-isoquinolinones, iodo-nitro-isoquinolinones and their homologues.

In another preferred embodiment of the invention the pADPRT-oxidizing ligands are selected from the group consisting of 3-nitroso-benzamide, 4-iodo-3-nitroso-benzamide, 2-iodo-5-nitroso-benzamide, 3-nitro-benzamide, 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide, 6-nitroso-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, 6-nitro-1,2-benzopyrone, and 5-iodo-6-nitro-1,2-benzopyrone.

In yet another preferred embodiment of the invention the pADPRT CCHC-oxidizing ligands are selected from the group consisting of 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide and 5-iodo-6-nitro-1,2-benzopyrone.

In still another preferred embodiment of the invention the non-covalent pADPRT-inhibitory ligands are selected from the group consisting of 6-amino-1,2-benzopyrone and 5-iodo-6-amino-1,2-benzopyrone.

In yet another preferred embodiment of the invention the GSH reducing agents are selected from the group consisting of diethylmaleate and BSO.

DESCRIPTION OF THE FIGURES

Figure 1 is a graph comparing the inactivation of pADPRT by NOBA and INOBA. pADPRT inactivation was performed in 20- μ l volumes of buffer containing 50 mM Hepes (pH 7.4), 100 mM KCl, 0.5 mM EDTA, pADPRT (0.4 μ g/ μ l) and different concentrations of inactivators. Inactivators were serially diluted in the above buffer from a 20 mM stock solution in DMF. The DMF concentration (2.5%) was kept constant in the inactivation mixes. After 2 hr. at 25°C, aliquots (3 μ l) were removed for assay of enzyme activity in a volume of assay mix (200 μ l) in which the inactivators were dilute enough not to act as inhibitors. The assay mix consisted of 100 mM Tris (pH 7.7), 14 mM 2-mercaptoethanol, 0.1 mM [32 P]-NAD $^{+}$, 0.2 mg/ml coDNA and 0.1 mg/ml histones. The result of

the pADPRT assays are shown in Figure 1. The incorporated ^{32}P , proportional to enzyme activity, is plotted as percent of the untreated sample.

5 Figure 2A is a graph comparing the effects of NOBA and INOBA on the enzyme activity and zinc content of pADPRT. Inactivation was performed as described in Example III. In these experiments $^{65}\text{Zn}^{2+}$ -labeled pADPRT was used. The Zn^{2+} content of pADPRT (upper panel) was determined as follows:
10 two parallels of aliquots (4 μl) were pipetted into 500 μl of ice-cold washing solution (50 mM Tris, pH 7.7, 10 mM 2-mercaptoethanol and 0.5 mM EDTA) then filtered onto GF/C disks presoaked in washing solution (cold), quickly washed 4 times with 1 ml of ice-cold washing solution, dried, and the
15 radioactivity determined. The protein-bound $^{65}\text{Zn}^{2+}$ is shown as percent of the untreated sample. Enzyme assays (lower panel) were performed as in Fig. 1.

Figure 2B shows the release of pADPRT zinc effected by
20 NOBA and INOBA. For the release of Zn^{2+} from transblotted pADPRT, the enzyme (3 μg per lane) was separated on SDS-PAGE, transblotted onto a nitrocellulose membrane, and renatured in the presence of $^{65}\text{ZnCl}_2$. The Zn^{2+} -loaded transblot was cut into strips and incubated with 2 ml of 50 mM Hepes/NaOH (pH
25 7.0) and 0.5 mM EDTA containing different concentrations of either NOBA or INOBA as indicated, for 2 hr. at 25°C. After incubation, 1 ml of the supernatant was counted for radioactivity, and the strips were washed 3 times with 2 ml of 50 mM Tris-HCl pH 7.4), 0.5 mM EDTA and 10 mM 2-
30 mercaptoethanol, and then dried and exposed to X-ray film. The intensity of the autoradiographic spots was quantitated by a scanner.

Figure 3 shows the time course of cell killings by
35 4-iodo-3-nitrobenzoic acid, INO_2BA and NOBA, respectively, alone and in combination with BSO. Molt 4-cells were seeded at a density of 0.05×10^6 cell/ml in the recommended growth

medium in the absence or the presence of drugs as indicated. Cells were incubated at 37°C in a 5% CO₂ atmosphere and proliferation was monitored in 14 hr. by cell counting in a hemacytometer. O, untreated control; ●, treated with 4 μM 4-iodo-3-nitrobenzoic acid; ▽, treated with 1 mM BSO; ▽, treated with 4 μM-iodo-3-nitrobenzoic acid and 1 mM BSO; □, treated with 4 μM 4-INO₂BA; ■, treated with 1 mM BSO; ▽, treated with 4 μM INO₂BA and 1 mM BSO. L-1210 cells were seeded at a density of 0.05 x 10⁶ cells/ml either from untreated stock suspensions or from 24-hr. pretreated (0.2 x 10⁶ cells/ml) suspensions. O, untreated control; ●, treated with 5 μM NOBA; ▽, treated with 1 mM BSO; ▽, 24-hr pretreatment with 1 mM BSO then treated with 4 μM NOBA.

Figure 4 shows the effects of INO₂BA alone (Figure 4A) and in combination (Figure 4B) with various concentrations of BSO on the viability of CEM x174 cells infected with SIV as determined by the tetrazolium assay. In both (A) and (B) the ordinate represents cell viability on day 10 of the assay and the abscissa represents the concentration of INO₂BA in the absence of BSO (Figure 4A), and in the presence of BSO (Figure 4B).

Figure 5 shows the effect of INO₂BA alone and in combination with BSO on SIV replication. Infected cell cultures described in Figure 4 were used for determination of supernatant virus titers by SIV p27 core antigen assay as described in Example XIV. The ordinate represents p27 ELISA performed on day 10 of the assay and the abscissa represents the concentration of INO₂BA alone (rear rows in panels A, B and C) and in combination with BSO (front rows in panels A, B and C). Panel D shows that various concentrations of BSO alone have no effect on SIV replication.

Figure 6 shows the effect of INO₂BA on SIVmac replication in CEM x-174 cells pre-infected with virus prior to treatment with INO₂BA, as determined by the reverse transcription assay.

Figure 7A shows the HIV-1 inactivation assay using NOBA and NOBP (6-nitroso-1,2-benzopyrone). The HIV-1 stock (HIV-1 100,000 TCID₅₀) was treated for 30 min. with 100 μ M NOBP at 22°C, the mixture was serial 10-fold diluted and inoculated 5 into PBL cultures. After 9 days the culture supernatants were harvested and the frequency of infected cultures was measured by immunoassay. The percent positive of cultures was then plotted as a function of the virus input titer. The relative amount of infectious virus available to cause 50% 10 infected cultures was decreased by 4 log units with NOBA.

Figure 7B shows an HIV-1 inactivation assay with NOBA at different temperatures. The assay was performed as described in Figure 7A except that the 30 min. preincubation of virus 15 with NOBA was carried out at 0°, 22° or 37°C.

Figure 7C shows the dose-response effect of NOBA on the viability of PHA-PBL. Cells (10⁶/ml) were treated with increasing doses of NOBA for 24 hours in the presence of MTT 20 indicator and the relative absorbance at 550 nm reflects the metabolic activity of the cells. The level of product formation in the absence of NOBA was considered to be 100% and all experimental values were normalized to that control value.

25

Figure 8 shows the effect of IABP on SIVmac239 replication. CEM x174 cells (3 x 10⁵/ml) were infected with SIVmac239 on day 0 and incubated for 3 days (37°C, 5% CO₂) before treatment with IABP (0-0.5 mM). The cultures were 30 incubated for an additional 4 days before being replenished with fresh CEM x174 cells (3 x 10⁵/ml) and IABP. On day 10 the levels of free virus in the supernatants of the cultures were assessed using the SIV p27 core antigen assay (Coulter Corp). The absorbance was measured at 450 nm.

35

Figure 9A shows the synergistic effect of IABP in combination with INO₂BA on SIVmac239 replication. Cells (1 x

10⁶/ml) were infected with SIVmac239 on day 0 and incubated for 3 days (37°C, 5% CO₂) before treatment with INO₂BA (0-300 μM) (rear columns) or with INO₂BA and IABP (0.3 mM) (front columns). The cultures were incubated for an additional 4 days before being replenished with fresh CEM x174 cells (3 x 10⁵/ml) and drug(s). On day 10 the levels of free virus in the supernatants were assessed using the SIV p27 core antigen assay, with absorbance measured at 450 nm.

10 Figure 9B shows the synergistic effect of IABP in combination with INO₂BA on the viability of SIVmac239-infected CEM x174 cells. The procedure was the same as for Figure 3A. The cell viability was measured by the MTT assay on day 10 of the experiment. The effect of INO₂BA alone is shown in the front columns, and the two drugs in combination in the rear columns.

Figure 10 shows the synergistic effect of IABP and INO₂BA, each alone and in combination, on the growth of L 1210 cancer cells.

DESCRIPTION OF SPECIFIC EMBODIMENTS

Definitions

25 As used herein:

"Biological Material" refers to any biological material extracted from a living organism, including blood, plasma, cerebrospinal fluid, organs, and the like, as well as the processed products of biological material extracted from a living organism.

"Zinc Finger" refers to a structural domain of a protein capable of binding a zinc atom. The nature of zinc finger protein domains is well described in the literature, e.g., Klug and Rhodes, Trends in Biochemical Sciences 12:464-469 (1987).

"Pharmaceutically acceptable acid addition salt" refers to those salts which retain the biological effectiveness and properties of the free bases and which are obtained by reaction with inorganic acids such as hydrochloric acid, 5 hydrobromic acid, sulfuric acid, nitric acid, phosphoric acid, methanesulfonic acid, ethanesulfonic acid, p-toluenesulfonic acid, salicylic acid and the like.

"pADPRT" refers to adenosinediphosphoribose transferase 10 also known as poly (ADP-ribose)polymerase, (EC 2.4.30), a specific DNA-binding nuclear protein of eucaryotes that catalyzes the polymerization of ADP-ribose. The enzymatic process is dependent on DNA.

15 "pADPRT CCHC-oxidizing ligands" refers to aromatic C-nitroso compounds that bind to the CCHC zinc finger of pADPRT and effect zinc ejection. pADPRT CCHC-oxidizing ligands also refers to C-nitro precursors of active aromatic C-nitroso compounds. Such ligands include, but are not limited to, 20 nitro-estradiol, iodo-nitro-estradiol, nitroso-estradiol, iodo-nitroso-estradiol, nitro-estrone, iodo-nitro-estrone, nitroso-estrone, iodo-nitroso-estrone, nitro-estriol, iodo-nitro-estriol, nitroso-estriol, iodo-nitroso-estriol, nitro-equilenin, iodo-nitro-equilenin, nitroso-equilenin, iodo- 25 nitroso-equilenin, nitro-equilin, iodo-nitro-equilin, nitroso-equilin, iodo-nitroso-equilin, nitro-diethylstilbestrol, iodo-nitro-diethylstilbestrol, nitroso-diethylstilbestrol, iodo-nitroso-diethylstilbestrol, nitrobenzamides, nitrosobenzamides, iodo-nitro-benzamides, 30 iodo-nitrosobenzamides, 4-iodo-3-nitrobenzamide, 4-iodo-3-nitrosobenzamide, 2-iodo-5-nitrobenzamide, 2-iodo-5-nitrosobenzamide, nitro-1,2-benzopyrones, nitroso-1,2-benzopyrones, 6-nitro-1,2-benzopyrones, 6-nitroso-1,2-benzopyrones, iodo-nitro-1,2-benzopyrones, iodo-nitroso-1,2- 35 benzopyrones, 4-iodo-3-nitro-1,2-benzopyrone, 4-iodo-3-nitroso-1,2-benzopyrone, 5-iodo-6-nitro-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, nitro-isoquinolinones, iodo-

nitro-isoquinolinones, nitroso-isoquinolinones, iodo-nitroso-isoquinolinones, 5-nitro-1(2H)-isoquinolinone, iodo-5-nitro-1(2H)-isoquinolinones, 5-nitroso-1(2H)-isoquinolinone, iodo-5-nitroso-1(2H)-isoquinolinones, 7-nitro-1(2H)-isoquinolinone, iodo-7-nitro-1(2H)-isoquinolinone, 7-nitroso-1(2H)-isoquinolinone, iodo-7-nitroso-1(2H)-isoquinolinones, 8-nitro-1(2H)-isoquinolinone, iodo-8-nitro-1(2H)-isoquinolinone, 8-nitroso-1(2H)-isoquinolinone, iodo-8-nitroso-1(2H)-isoquinolinones and their homologues.

10

"GSH Decreasing Agent" refers to agents that decrease cellular levels of glutathione. Examples include, but are not limited to diethylmaleate and BSO.

15

"Non-covalent pADPRT-inhibitory ligands" refers to compounds that bind non-covalently to the nicotinamide site of pADPRT, but that do not attack the F1 zinc finger of pADPRT oxidatively. Examples of such ligands include, but are not limited to, 1,2-benzopyrones, benzamides, amino-1,2-benzopyrones, amino-benzamides, 6-amino-1,2-benzopyrones, 3-amino-benzamide, amino-iodo-benzopyrones, amino-iodo-benzamides, 5-iodo-6-amino-1,2-benzopyrone, 4-iodo-3-amino-benzopyrone and their homologues.

20

25 "Alkyl" refers to saturated or unsaturated branched or straight chain hydrocarbon radical. Typical alkyl groups includes methyl, ethyl, propyl, isopropyl, butyl, isobutyl, tertiary butyl, pentyl, hexyl and the like.

30

"Alkoxy" refers to the radical -O-alkyl. Typical alkoxy radicals are methoxy, ethoxy, propoxy, butoxy and pentoxy and the like.

"Cycloalkyl" refers to a saturated monocyclic hydrocarbon radical containing 3-8 carbon atoms such as cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl and cyclooctyl.

"Substituted phenyl" refers to all possible isomeric phenyl radicals mono or di-substituted with a substituent selected from the group consisting of alkyl, alkoxy, hydroxy or iodo.

5

The subject invention provides for novel synergistic compositions that find use as anti-tumor or anti-viral compositions. In one embodiment of the invention, the synergistic compositions comprise one or a plurality of pADPRT CCHC-oxidizing ligands in combination with one or a plurality of agents selected from the group consisting of agents that decrease cellular levels of glutathione and non-covalent pADPRT-inhibitory ligands.

15

In another embodiment of the invention the synergistic compositions comprise one or a plurality of pADPRT CCHC-oxidizing ligands in combination with one or a plurality of GSH decreasing agents.

20

In yet another embodiment of the invention the synergistic compositions comprise one or a plurality of pADPRT CCHC-oxidizing ligands in combination with one or a plurality of non-covalent pADPRT-inhibitory ligands.

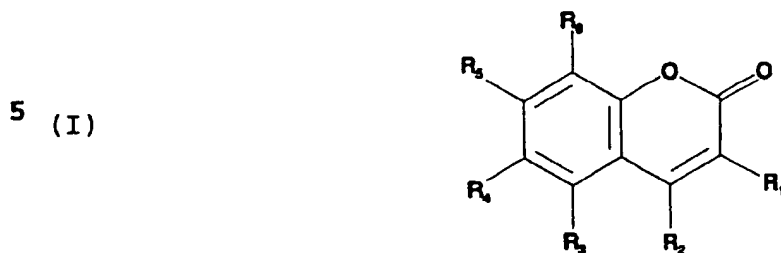
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The pADPRT CCHC-oxidizing ligands of the subject invention are described by the formulae below:

30

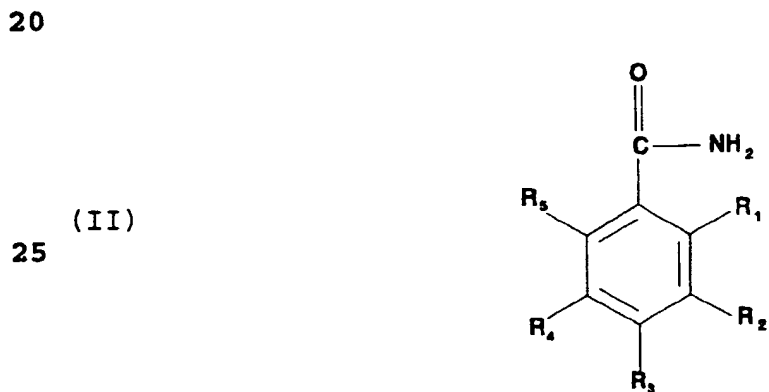
35

Compound I has the following formula:



10 wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro.

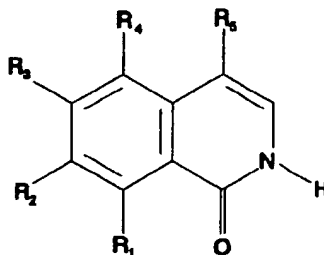
Compound II has the formula:



30 wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or nitro.

Compound III has the formula:

5 (III)



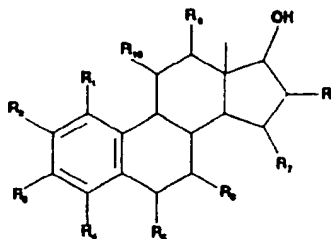
10 wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of
15 the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or nitro.

Compound IV has the formula:

20

(IV)

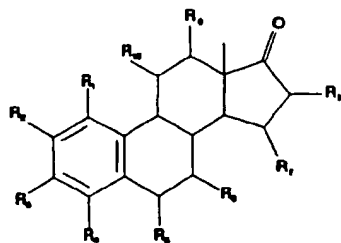
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30 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10}
35 substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro.

Compound V has the formula:

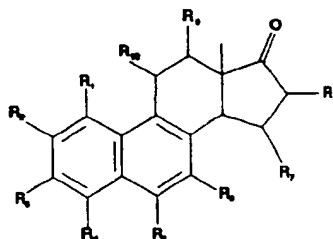
5 (V)



10 wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ are,
independent of one another, selected from the group
consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆)
alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and
pharmaceutically acceptable salts thereof, wherein at least
15 three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀
substituents are hydrogen and wherein at least one of the ten
R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso
or nitro.

20 Compound VI has the formula:

(VI)

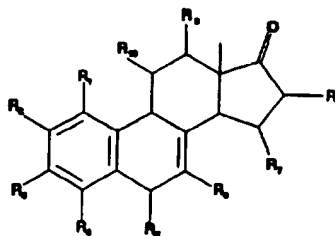


25

wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ are,
independent of one another, selected from the group
30 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆)
alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and
pharmaceutically acceptable salts thereof, wherein at least
three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀
substituents are hydrogen and wherein at least one of the ten
35 R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso
or nitro.

Compound VII has the formula:

5 (VII)

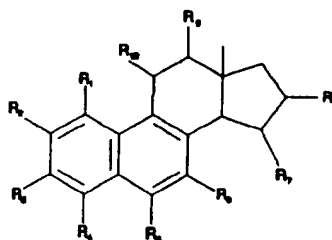


wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ are,
 10 independent of one another, selected from the group
 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆)
 alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and
 pharmaceutically acceptable salts thereof, wherein at least
 15 three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀
 substituents are hydrogen and wherein at least one of the ten
 R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso
 or nitro.

Compound VIII has the formula:

20

(VIII)

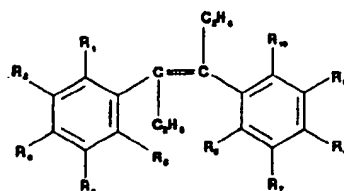


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wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ are,
 independent of one another, selected from the group
 30 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆)
 alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and
 pharmaceutically acceptable salts thereof, wherein at least
 three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀
 substituents are hydrogen and wherein at least one of the ten
 35 R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso
 or nitro.

Compound IX has the formula:

5 (IX)



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are,
 10 independent of one another, selected from the group
 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6)
 alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and
 pharmaceutically acceptable salts thereof, wherein at least
 three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10}
 15 substituents are hydrogen and wherein at least one of the ten
 R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso
 or nitro.

In a preferred preferred embodiment, compounds of
 20 formulae I-IX have an iodo group attached adjacent to a
 nitroso or nitro group.

In a more preferred embodiment, compounds of formulae
 I-IX have an iodo group attached adjacent to a nitro group.

25

In another preferred embodiment, the pADPRT CCHC-
 oxidizing ligands are selected from the group consisting of a
 compound according to formula I and a compound according to
 formula II.

30

In another preferred embodiment the pADPRT CCHC-
 oxidizing ligands are selected from the group consisting of
 3-nitroso-benzamide, 4-iodo-3-nitroso-benzamide, 2-iodo-5-
 nitroso-benzamide, 3-nitro-benzamide, 4-iodo-3-nitro-
 35 benzamide, 2-iodo-5-nitro-benzamide, 6-nitroso-1,2-
 benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, 6-nitro-1,2-
 benzopyrone, and 5-iodo-6-nitro-1,2-benzopyrone.

In another preferred embodiment the pADPRT CCHC-oxidizing ligands are selected from the group consisting of 4-iodo-3-nitrobenzamide, 2-iodo-5-nitrobenzamide and 5-iodo-6-nitro-benzamide-1,2-benzopyrone.

5

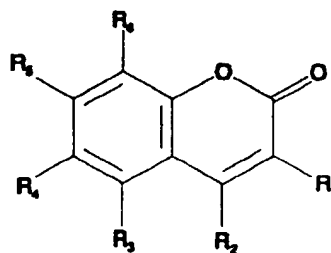
Non-covalent pADPRT-inhibitory ligands of the subject invention are described by the formulae below:

Compound X has the formula:

10

(X)

15



20

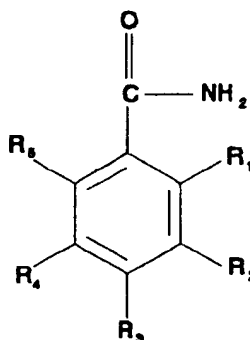
wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, amino, iodo, alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen.

25

Compound XI has the formula:

30

(XI)



35

wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen,

hydroxy, amino, iodo, alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R₁, R₂, R₃, R₄, R₅ and R₆ substituents are hydrogen.

5

In a preferred embodiment of the invention the non-covalent pADPRT-inhibitory ligands are selected from the group consisting of 6-amino-1,2-benzopyrone and 5-iodo-6-amino-1,2-benzopyrone.

10

GSH decreasing agents of the subject invention include, but are not limited to, BSO and diethylmaleate. In a preferred embodiment said GSH decreasing agents are selected from the group consisting of diethylmaleate and BSO.

15

Synergistic Anti-Tumor Activity of the Novel Compositions

pADPRT CCHC-oxidizing ligands such as aromatic C-nitroso compounds are potent anti-tumorigenic drugs. The anti-tumorigenic activity of such compounds has been studied extensively and is described in the literature and in Example II. Aromatic C-nitro compounds are in vivo precursors of aromatic C-nitroso compounds. These in vivo precursors are reduced to the active C-nitroso ligand in vivo, supplying a readily available source of the active C-nitroso ligand in vivo. Such activity has been studied extensively in cell lines and is described in detail in Example II.

Example VI describes the metabolic conversion of INO₂BA to 4-iodo-3-aminobenzamide ("INH₂BA") by 855-2 cells. A slow but steady reduction of the nitro compounds occurred at a rate of 2.2±0.2% per 18 hours of cell incubation at 37°C. Apart from trace metabolites, the only identifiable metabolite was 4-iodo-3-amino-benzamide (INH₂BA).

Example III illustrates the inactivation of pADPRT by NOBA and INOBA, with concomitant zinc ion ejection. As shown in the example, under comparable in vitro conditions INOBA was the more effective inactivator of pADPRT. Zinc ejection was complete within 2 hours for both NOBA and INOBA.

Example IX illustrates the synergistic effect of BSO in combination with INO_2BA in various cancer cell lines. BSO in combination with INO_2BA showed synergistic cytotoxic effects in all twelve of the cell lines tested.

5 Example VIII illustrates the synergistic effect of BSO with 4-iodo-3-nitrobenzoic acid, INO_2BA and NOBA in Molt-4 and L-1210 cells. As shown in the example, treatment of Molt-4 and L-1210 cells with 1 mM BSO alone had no effect on cell growth over two days. Treatment of cells with 4-iodo-3-
10 nitrobenzoic acid, INO_2BA and NOBA alone also had no inhibitory effect on cell growth over two to three days. However, treatment with BSO in combination with the pADPRT CCHC-oxidizing ligands synergistically produced cell death of tumor cells.

15 Thus, as is demonstrated by the above, the exemplary compositions of BSO in combination with 4-iodo-3-nitrobenzoic acid, INO_2BA and NOBA, respectively, resulted in synergistic cytotoxic effects in various cancer cell lines. Specifically, BSO increased the cytotoxicity of each of NOBA,
20 INO_2BA and 4-iodo-3-nitrobenzoic acid. The increased cytotoxicity in each case was more than additive.

Example XVII demonstrates the synergistic effect of IABP in combination with INO_2BA in L 1210 cells.

Thus, as is demonstrated by the above, the exemplary
25 composition of IABP in combination with INO_2BA resulted in synergistic cytotoxic effects in L 1210 cells. Specifically, INO_2BA increased the cytotoxicity of IABP and IABP increased the cytotoxicity of INO_2BA . The increased cytotoxicity in each case was more than additive.

30

Synergistic Anti-Retroviral Activity of the Novel Compositions

pADPRT CCHC-oxidizing ligands are also potent anti-retroviral drugs. Such activity has been studied extensively
35 and is described in Example XIII. Aromatic C-nitro compounds are in vivo precursors of active C-nitroso derivatives. These precursors are reduced in vivo to the active C-nitroso

compound, providing a ready supply of the active C-nitroso compound in vivo. The anti-viral activity of such C-nitro precursors has been studied and is described in Example XI.

While the C-nitro pro-drug had no effect on SIV infectivity, its intra-cellular metabolite readily inhibits SIV propagation. A further advantage of the pro-drug is that it is effective even after viral infection, whereas the C-nitroso drug requires pre-incubation of cells prior to viral infection, as is demonstrated in Example XV.

10 Example XIV illustrates the effect of various concentrations of INO_2BA alone and in combination with various concentrations of BSO on the viability of SIV-infected CEM x174 cells.

Example XIV also shows the effect of various
15 concentrations of INO_2BA alone and in combination with various concentrations of BSO on SIV replication. As is demonstrated in the Example, BSO alone had no effect on SIV replication. A BSO concentration of 0.01 mM in combination with 50 μM and 100 μM INO_2BA showed a synergistic effect on SIV replication
20 activity. Both 0.05 and 0.25 mM BSO in combination with 25 μM , 50 μM and 100 μM INO_2BA showed a synergistic effect on SIV replication activity.

Thus, as is demonstrated by the above, the exemplary composition of BSO in combination with INO_2BA resulted in a
25 potentiation effect on SIV replication.

Non-covalent pADPRT-inhibitory ligand IABP blocked the infectious cycle of HIV as tested in various cell lines. Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991).

30 The molecular mode of action of pADPRT CCHC-oxidizing ligands is clearly different from that of non-covalent pADPRT-inhibitory ligands. See Rice et al., Nature 361:473-475 (1993); Chuang et al., FEBS Lett. 326:140-144 (1993); Rice et al., Proc. Natl. Acad. Sci. 90:9721-9724
35 (1993); and Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991). That the modes of action of pADPRT CCHC-oxidizing ligands and non-covalent pADPRT ligands are

different is illustrated in Example XIII. Example XIII shows that depletion of cellular GSH by BSO, an inhibitor of γ -glutamyl cysteine synthetase, Meister, Science 220:472-477 (1991), synergistically enhances the inhibitory effect of
5 INOBA and INO₂BA on SIV replication in CEM x174 cells. Since IABP was insensitive to addition of BSO, Chuang et al., submitted (1994), the molecular modes of action of the pADPRT CCHC-oxidizing ligands and non-covalent pADPRT ligands are clearly differentiated.

10 Example XVIII shows the inhibitory effect of IABP on SIVmac239 replication. As is clear from the example, both 0.3 and 0.5 mM IABP reduced the titre of virions in the supernatant of the cultures.

The synergistic effect of the pADPRT CCHC-oxidizing
15 ligand INO₂BA in combination with the non-covalent pADPRT ligand IABP on the replication of SIV is illustrated in Example XIX. As is shown in the Example, a more than additive anti-HIV action of 0.3 mM IABP and varying concentrations of INO₂BA, as assayed by the inhibition of p27
20 formation in CEM x174 cells infected with SIVmac239, was observed. This drug combination also supported cell growth more than additively.

The non-toxicity of the non-covalent pADPRT-inhibitory ligand IABP is shown in Example XX.

25

Administration

In carrying out the methods of the present invention, the synergistic compositions will be administered in amounts which will be sufficient to kill cancer cells or inhibit
30 retroviral expression or infection in the host in the pharmaceutical form most suitable for such purposes.

Administration of the synergistic compositions and salts described herein can be via any of the accepted modes of administration for therapeutic agents. These methods include
35 systemic or local administration such as oral, parenteral, transdermal, subcutaneous, or topical administration modes. The preferred method of administration of these compositions

is intravenous, except in those cases where the subject has topical tumors or lesions, where the topical administration may be proper. In other instances, it may be necessary to administer the compositions in other parenteral or even oral
5 forms.

Depending on the intended mode, the compositions may be in the solid, semi-solid or liquid dosage form, such as, for example, injectables, tablets, suppositories, pills, time-release capsules, powders, liquids, suspensions, or the like,
10 preferably in unit dosages. In one embodiment the compositions will include an effective amount of one or a plurality of pADPRT CCHC-oxidizing ligands of formulae I-IX, or the pharmaceutically acceptable salts thereof, in combination with one or a plurality of GSH decreasing agents
15 and in addition, it may include any conventional pharmaceutical excipients and other medicinal or pharmaceutical drugs or agents, carriers, adjuvants, diluents, etc., as is customary in the pharmaceutical sciences.

20 In another embodiment the compositions will include an effective amount of one or a plurality of pADPRT CCHC-oxidizing ligands of formulae I-IX, or the pharmaceutically acceptable salts thereof, in combination with one or a plurality of non-covalent pADPRT-inhibitory ligands of
25 formulae X-XI, or the pharmaceutically acceptable salts thereof, and in addition it may include any conventional pharmaceutical excipients and other medicinal or pharmaceutical drugs or agents, carriers, adjuvants, diluents, etc., as is customary in the pharmaceutical
30 sciences.

For solid compositions such excipients include pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like may be used. The
35 synergistic pharmaceutical compositions may be also formulated as suppositories using, for example, polyalkylene glycols, for example, propylene glycol, as the carrier.

Liquid, particularly injectable pharmaceutical compositions can, for example, be prepared by dissolving, dispersing, etc., an effective amount of one or a plurality of pADPRT CCHC-oxidizing ligands of formulae I-IX, or the
5 pharmaceutically acceptable salt thereof, in combination with an effective amount of one or a plurality of GSH decreasing agents or in combination with an effective amount of one or a plurality of non-covalent pADPRT-inhibitory ligands, or the pharmaceutically acceptable salt thereof, in a pharmaceutical
10 solution such as, for example, water, saline, aqueous dextrose, glycerol, ethanol, and the like, to thereby form the injectable solution or suspension.

If desired, the pharmaceutical composition to be administered may also contain minor amounts of nontoxic
15 auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and the other substances such as for example, sodium acetate, triethanolamine oleate, etc.

If desired, the pharmaceutical composition to be administered may contain liposomal formulations comprising a
20 phospholipid, a negatively charged phospholipid and a compound selected from cholesterol, a fatty acid ester of cholesterol or an unsaturated fatty acid. The synergistic composition may be encapsulated or partitioned in a bilayer of liposomes of the liposomal formulation according to U.S.
25 Patent Application No. 08/020,035 entitled "Liposomal Formulations and Methods of Making and Using Same" filed on February 19, 1993 which is incorporated herein by reference.

Parenteral injectable administration is generally used for subcutaneous, intramuscular or intravenous injections and
30 infusions. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions or solid forms suitable for dissolving in liquid prior to injection.

A more recently devised approach for parenteral administration employs the implantation of a slow-release or
35 sustained-released systems, which assures that a constant level of dosage is maintained, according to U.S. Patent No. 3,710,795, which is incorporated herein by reference.

In one embodiment, such pharmaceutical generally compositions include about 1 to about 100 parts pADPRT CCHC-oxidizing ligands of formulae I-IX, or the pharmaceutically acceptable salts thereof, and about 200 to about 800 parts 5 GSH decreasing agents mixed with a pharmaceutical excipient.

In one embodiment such pharmaceutical compositions include about 1 to about 100 parts pADPRT CCHC-oxidizing ligands of formulae I-IX, or the pharmaceutically acceptable salts thereof, and about 200 to about 800 parts non-covalent 10 pADPRT-inhibitory ligands of formulae X-XI, or the pharmaceutically acceptable salts thereof, mixed with a pharmaceutical excipient.

Actual methods of preparing such dosage forms are known, or will be apparent to those skilled in this art, and are 15 desired in detail described in Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pennsylvania, 17th Edition, 1985. The composition or formulation to be administered will, in any event, contain such quantity of the synergistic compositions that will assure that a 20 therapeutically effective amount will be delivered to a patient. A therapeutically effective amount means an amount effective to prevent development of or to alleviate the existing symptoms of the subject being treated.

The amount of synergistic composition administered will, 25 of course, be dependent on the subject being treated, on the subject's weight, the severity of the affliction, the manner of administration and the judgment of the prescribing physician. However, an effective dosage is one wherein one or a plurality of pADPRT CCHC-oxidizing ligands are 30 administered in the range of 0.001 to 5000 mg/kg/day, preferably 0.01 to 1000 mg/kg/day, and more preferably 0.1 to 100 mg/kg/day.

Although the effective molar ratio of pADPRT CCHC-oxidizing ligands to GSH decreasing agents or to non-covalent 35 pADPRT-inhibitory ligands comprising such synergistic compositions will vary in accordance with the nature of pADPRT CCHC-oxidizing ligands, non-covalent pADPRT-inhibitory

ligands, GSH decreasing agents and particular subject, preferred molar ratios of pADPRT CCHC-oxidizing ligands to GSH decreasing agents are from about 1:1 to about 1:10, and more preferably from about 1:1 to about 1:100. Preferred
5 effective molar ratios of pADPRT CCHC-oxidizing ligands to non-covalent pADPRT-inhibitory ligands are from about 1:1 to about 1:10, and more preferable from about 1:1 to about 1:100. Effective molar ratios may be readily determined by testing the effect of a range of concentrations of one or a
10 plurality of C-nitroso pADPRT CCHC-oxidizing ligands in combination with a range of concentrations of GSH decreasing agents or non-covalent pADPRT-inhibitory ligands.

Generally, the upper limit for the drug dose determination is its efficacy balanced with its possible
15 toxicity. Such toxicity appears to be very low for the pADPRT CCHC-oxidizing ligand prodrug INO₂BA (hamsters injected daily intraperitoneally with 200 mg/kg for 12 days showed no toxic effects), for the non-covalent pADPRT-inhibitory ligand IABP, Cole *et al.*, Biochem. Biophys. Res. Commun. **180**:504-514
20 (1991), and for BSO, Liebman *et al.*, Cancer Res. **53**:2066-2070 (1993); Yao *et al.*, Cancer Res. **53**:3662-3666 (1993); Terradez *et al.*, Biochem. J. **292**:477-483 (1993); and Polhuijs *et al.*, Biochem. J. **285**:401-404 (1992). Thus, the administered dose may be as high as needed to achieve desirable therapeutic
25 effect.

Various substituents of the ligands as shown in formulae I-XI, are likely to modify lipid solubility or rate of cellular penetration, thus clinical dosage schedules of the synergistic compositions is likely to be altered on a
30 molecular level by added substituents.

The chemotherapy may be repeated intermittently while tumors or HIV infections are detectable or even when they are not detectable.

Moreover, due to its apparent nontoxicity, the therapy
35 may be provided alone or in combination with other antiviral or other drugs, such as for example AZT, anti-inflammatories antibiotics, corticosteroids, vitamins and other such drugs.

There are no contraindications to use the synergistic compositions with even such toxic drugs as AZT since the synergistic compositions are nontoxic and their modes of action are quite different. Possible synergism between the 5 synergistic compositions and other drugs is expected and predictable. In addition, possible synergism between a plurality of synergistic compositions of the present invention is also expected and predictable.

The synergistic compositions are equally useful for 10 treatment of herpetic lesions caused by both HSV-1 and HSV-2. The compositions would be preferably administered by intravenous infusion or other parenteral or systemic mode of administration. In case of sores, the synergistic composition could also be administered topically. Infection 15 caused by CMV would be treated preferably in the same fashion as that suggested for AIDS treatment.

The subject invention also provides for methods of decreasing the titer of infectious viruses, particularly 20 retroviruses (including the retrovirus HIV-1) in biological materials by inactivating the viruses with one or a plurality of synergistic compositions (Fig. 8). Viruses may be inactivated by contact between a synergistic composition of interest and the virus. The term "decreasing" includes the 25 complete elimination of all the infectious viruses of interest, as well as a diminution in the titer of the infectious viruses. It is of particular interest to decrease the number of infectious viruses in biological material that is to be introduced into a living organism so as to reduce 30 the possibility for infection. It is also of interest to decrease the titer of infectious viruses that might be present in or on non-biological materials that come into contact with living organisms. Such non-biological materials include surgical instruments, dental instruments, hypodermic 35 needles, public sanitary facilities, and the like. Inactivation of HIV virions occurs by direct acting C-nitroso

pADPRT CCHC-oxidizing ligands only-- not by C-nitro precursors.

The subject methods inactivate viruses by employing the step of contacting an effective amount of one or a plurality of synergistic compositions to the biological or non-biological material of interest. A preferred embodiment of the subject method is decreasing the concentration of infectious virus in blood. Another preferred embodiment of the subject invention is the inactivation of AZT resistant viruses and retroviruses, particularly HIV and SIV.

The subject invention also provides for methods of inducing apoptosis in a tumor cell by administering one or a plurality of synergistic compositions to a cell culture or a mammalian host having one or a plurality of tumors.

The invention having been described, the following examples are offered to illustrate the subject invention by way of illustration, not by way of limitation.

20

EXAMPLES

I. Syntheses

25 A. 6-Nitroso-1,2-Benzopyrone

An example of a method for the preparation of 6-nitroso-1,2-benzopyrone is provided as follows:

To a stirred solution of 6-amino-1,2-benzopyrone hydrochloride (4.00 g, 20 mmol) in water (40 ml) at 22°C was added a solution of sodium tungstate (5.93 g, 20 mmol) in water (20 ml) followed by 30% aqueous hydrogen peroxide (5 ml) and stirring was continued for 1.5 hours. The oxidation product was extracted from the green-colored mixture with two 100 ml volumes of ethyl acetate, the combined extracts washed with 0.1 N HCl (50 ml) and then water (100 ml). The ethyl

acetate was removed by rotary evaporation and the residue recrystallized from warm ethanol (250 ml).

Analysis of Reaction Product

5 The green crystals obtained from the recrystallization step (1.48 g, 42% yield) displayed light absorption at 750 nm characteristic of monomeric arylnitroso compounds. Mass spectrum: m/z (relative intensity): 175 (M^+ , 100), 161 (16.88), 145 (33.77), 133 (10.38), 117 (56.09), 89 (79.71), 10 63 (57.13). High resolution data for the M^+ peak: calculated for C_9H_5NO ; 175.0268; found: 175.0271 (deviation = 1.1 ppm). 1H -NMR ($CDCl_3$, 300 MHz) δ (ppm) from TMS: doublet (6.572 and 6.604) H-4 split by H-3; doublet (7.472 and 7.501) H-8 split by H-7; doublet of doublets (7.860/7.866 and 7.889/7.798) H-7 15 split by H-8 and finely split by H-5; doublet (7.910 and 7.942) H-3 split by H-4; doublet (8.308 and 8.315) H-5 finely split by H-7. UV/VIS spectrum in ethanol, λ max (ϵ): 750 nm (46), 316 nm (8.96×10^3), 274 nm (2.24×10^4). Melting Point: The compound polymerizes above 160°C, blackens and 20 melts in the range of 325-340°C.

This nitroso-compound may also be prepared by reacting 6-amino-1, 2-benzopyrone (as the free base) with 3-chloroperoxybenzoic acid in ethyl acetate or halocarbon solvents.

25

B. 3-nitrosobenzamide, 2-nitrosobenzamide and 4-nitrosobenzamide

To a stirred solution of 3-aminobenzamide (Aldrich Chemical Co.) (0.476 g, 3.50 mmol) in ethyl acetate (50 ml) 30 at ambient temperature was added 1.208 g of 3-chloroperoxybenzoic acid (commercial grade, 50-60% purity, Aldrich), whereupon the solution turned green. After 10 minutes the mixture was extracted with 0.14 M aqueous sodium bicarbonate (58 ml), washed with three successive 40-ml 35 portions of water, dried over sodium sulfate, then reduced in volume to 20 mL by rotary evaporation and placed in the freezer (-20°C), whereupon the product slowly deposited as a

light yellow solid during a period of 72 hours (0.180 g, 34% yield).

The 2-nitrosobenzamide and 4-nitrosobenzamide isomers may be similarly prepared by oxidizing 2-aminobenzamide and 5 4-aminobenzamide, respectively.

Analysis of Reaction Product

Melting point: The substance darkens above 135°C, softens and apparently polymerizes in the range 150-160°C, 10 and melts at 240-250°C (with decomposition). In solution the compound is green-blue. Mass spectrum: m/z (relative intensity): 150 (M⁺, 100), 136 (10.9), 120 (77.2), 103 (31.6), 92 (46.5), 85 (22.8), 71 (33.3). High resolution data for the M⁺ peak: calculated for C₇H₆N₂O₂: 150.04292 found: 15 150.042900 (deviation = 0.2 ppm). NMR spectrum: ¹H-NMR (DMSO-d₆, 300 MHz) δ (ppm) from TMS: broad singlet (7.737) N-H; t (7.824, 7.850, 7.875) H-5 split by H-4 and H-6; d (8.059 and 8.086) H-6 split by H-5; d (8.357 and 8.383) H-4 split by H-5; s (8.472) H-2. The singlet at 7.737 corresponds to 1 20 proton; the second N-H proton, spectrally non-equivalent in this compound, is overlaid by the doublet of H-4. This doublet integrates to 2 protons and can be resolved by addition of D₂O to the DMSO solution. UV-VIS absorption spectrum in absolute ethanol, λ_{max} (ε): 750 nm (37.6), 304 nm 25 (5.35 x 10³) and 218 nm (1.50 x 10⁴). An absorption maximum at 750 nm is characteristic of monomeric aryl nitroso compounds.

In another embodiment, 3-nitrosobenzamide is synthesized by dissolving 3-aminobenzamide (5.0 g) in N,N- 30 dimethylformamide (DMF) solvent (25 ml) and then chilled in an ice bath. 3-Chloroperoxybenzoic acid (2.1 equivalents) is also dissolved in DMF solvent (25 ml) in a 250-ml flask equipped with a stirrer and thermometer and, as needed, an ice bath. This solution is chilled to 0-5°C, the ice bath is 35 removed, and to it, with stirring, is added all at once the chilled 3-aminobenzamide solution. The mixture immediately becomes a transparent brown color, but within 0.5 minute

turns to a deep green, and within 1.0 minute the temperature rises to 70°C, at which time the ice bath is reapplied to the reaction flask whereupon the temperature begins to fall, and is allowed to fall to 25°C, and stirring is continued for a total of 5 minutes. Some precipitation occurs (3,3'-azoxybenzamide side-product), thereafter the mixture is chilled to 5°C for 10 minutes. The chilled mixture is filtered (suction) to remove the azoxy precipitate, and the green filtrate is poured into chilled (5-10°C) and stirred aqueous 0.40 M Na₂CO₃ (200 ml), resulting in a light green suspension, and the suspension is stirred for an additional 10 minutes at 5-10°C to assure maximal product precipitation. Note that the pH of the suspension is about 8.5, which assures that 3-chlorobenzoic acid is retained in the aqueous solution as the sodium salt. The precipitate is then collected on a suction funnel and rinsed with deionized water (100 ml). This material, which is 3-NOBA (mostly as the tan dimer) containing residual 3,3'-azoxybenzamide side-product impurity, is then transferred, while damp, to a suitable flask and to it is added 50% aqueous acetic acid (200 ml). The mixture is warmed to 65-70°C to dissolve the dimer into the soluble monomeric 3-NOBA (green) and stirred for 5 minutes at 65°C. The azoxy impurity (yellow) is poorly soluble and remains undissolved. The warm mixture is filtered (gravity) to give a clear green filtrate, which is allowed to cool. It is then chilled and placed in the refrigerator freezer (-20°C) overnight to allow the 3-NOBA to redeposit as the light tan solid dimer. On the following day the solid product is collected on a suction filter, rinsed with fresh solvent, and the product cake is then dried by vacuum under mild warming for several hours. One typically obtains 2.24 g of dry 3-NOBA containing a trace of the azoxy impurity. The product is recrystallized by dissolving it again in 50% aqueous acetic acid (120 ml) and allowing to redeposit overnight in the freezer. After collection, rinsing and drying *in vacuo*, the weight is 2.08 g (37%

overall yield). TLC shows the material is 3-NOBA with a trace of the azoxy impurity.

C. 4-Iodo-3-Nitrobenzamide

5 In a 100 -mL flask equipped with a magnetic stirrer, thermometer and ice bath, a stirred solution of 4-Iodo-3-nitrobenzoic acid (1025 mg, 3.50 mMoles) (Chemica Alta Ltd., Edmonton, Alberta, Canada) in N,N-dimethylformamide (10 mL) is cooled to 10°C, and then thionyl chloride (0.76 mL, 10.5
10 mMoles) is added to it. There is no exothermicity, the ice bath is removed, and the solution is allowed to warm to ambient temperature, and stirring is continued for a total of 1 hour. Then the solution is poured into chilled, concentrated ammonium hydroxide (20 mL), resulting in a dark
15 yellow mixture, which is stirred for 5 minutes. Then chilled deionized water (50 mL) is added, causing precipitation of the light yellow product. After allowing the precipitation mixture to stand chilled on ice for 10 minutes, the precipitate is collected on a suction filter, rinsed with
20 cold water, and then dried by vacuum pumping. The resultant crude product (500.4 mg) is then re-crystallized by dissolving it in acetonitrile (7.0 mL) heated to about 65°C, followed by cooling and allowing the solution to stand in the refrigerator overnight. The yellow crystals are collected,
25 rinsed with chilled solvent and dried by vacuum pumping, to give 415.2 mg (40.5% yield) of 4-Iodo-3-nitrobenzamide, m.p. 152-155°C.

¹H NMR spectrum, in DMSO-d₆ δ (ppm) values relative to
30 TMS): broad singlet (7.67) due to one nonequivalent proton of the amido NH₂ group; doublet of doublets (7.84, 7.85 and 7.86, 7.87) due to H-5 split by H-6 and finely split by H-2; doublet (8.22, 8.24) due to H-6 split by H-5; broad singlet centered near 8.22, overlapping the signal of H-6, due to the
35 second nonequivalent proton of the amido NH₂ group; doublet (8.35, 8.36) due to H-2 finely split by H-5. At higher NMR field signals due to adventitious water (2.5 ppm),

deuterated-DMSO impurity protons (3.3 ppm) and crystallization solvent acetonitrile (single at 2.07 ppm) are observed. Integration of the acetonitrile signal indicates approximately one molecule of acetonitrile per 3 molecules of 5 4-iodo-3-nitrobenzamide.

UV absorption spectrum in absolute ethanol, λ max (ϵ): 308nm (1.59×10^3), 242nm (1.31×10^4), 208nm (1.45×10^4).

- 10 Elemental analysis (Schwarzkopf Microanalytical Laboratory): Calculated for $C_7H_5N_2O_3I$: C, 28.79%; H, 1.72; I, 43.46; N, 9.59. Found: C, 29.63; H, 1.72; I, 41.47; N, 9.99. Deviations from calculated are believed to be due to the presence of acetonitrile (crystallization solvent) as
15 detected in the NMR spectrum. High resolution EI mass spectrum: calculated for $C_7H_5N_2O_3I$: 291.9345; Found: $M^+(m/z)$: 291.9349 (deviation= -1.4 ppm).

D. 4-Iodo-3-Nitrosobenzamide

- 20 The method of synthesis of INOBA differs from that for 4-iodo-3-nitrosobenzamide in that a different oxidant is required. In a procedure favoring oxidation to the nitroso state, each of 20 test-tubes (13 x 100 mm) was charged with 5.2 mg (0.020 mmol) of INH_2BA (synthetic), Mendeleyev et al.
25 (1994), see also Example I.J., and 0.100 ml of absolute EtOH. Each tube was heated briefly to dissolve the amino compound, followed immediately by H_2O (0.400 ml) and then peracetic acid reagent (0.400 ml, 32% in dilute acetic acid) added at a fast drip during 10 sec. After 2 min., ethyl acetate (2.0 ml) and
30 H_2O (2.0 ml) were added, the mixture vortexed, and then stored on ice while the other tubes were identically processed. The upper (ethyl acetate) layers were pooled, the solution concentrated (rotary evaporation) to 4 ml, H_2O added until turbid, and placed at 2°C for 18 h. The product deposited as
35 a light tan solid. Yield: 35.3 mg (32%), m.p. 174-176°C (decomp). High resolution EIMS: calculated for $C_7H_5N_2O_2I$:

275.9396; found $M^+(m/z)$; 275.9399 (deviation = -0.3 ppm). 1H NMR (DMSO- d_6) δ (ppm): 6.65 (1H, d, $J=2.2$ Hz), 7.60 (1H, s), 8.02 (1H, dd, $J=8.1$ Hz, $J=2.2$ Hz), 8.18 (1H, s), 8.50 (1H, d, $J=2.2$ Hz). UV: max 370, 252, 226 nm and shoulders at 315, 293 nm. Vis: max 750 nm.

E. Nitroso-1(2H)-isoquinolinones (a mixture of 5-nitroso and 7-nitroso isomers)

1-(2H)-Isoquinolinone (isocarbostyryl) (Aldrich) was
10 nitrated using a general method for isoquinolinone compounds.

C.G. LeFevre and R.J. W. LeFevre, J. Chem. Soc. 1470 (1935). The nitration product (a mixture of the 5-nitro and 7-nitro isomers, as assigned by Y. Kawazoe and Y. Yoshioka, Chem Pharm. Bull. (Tokyo) 16:715-720 (1968), although one of the
15 isomers could be the 8-nitro isomer) was then reduced to the corresponding amino-1(2H)-isoquinolinones using a combination of potassium borohydride and palladium-on-carbon catalyst in aqueous methanol. To the resultant amino-1(2H)-isoquinolinones (as free bases) (0.560 g. 3.50 mmol) in ethyl
20 acetate (175 mL) at 30°C was added 1.208 g of 3-chloroperoxybenzoic acid (Aldrich). The mixture became cloudy and after 20 minutes it was filtered, extracted with 0.14M sodium bicarbonate (58 mL), washed with two 50-mL portions of water, and dried over sodium sulfate. The volume
25 of the solution was reduced to 50 mL by rotary evaporation and then placed in the freezer (-20°C), whereupon an orange solid product was deposited (0.102 g).

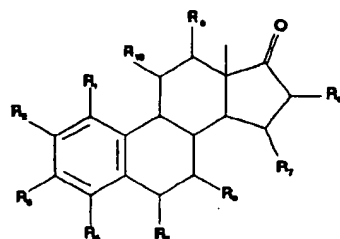
Analysis of Reaction Product

30 Melting point: substance darkens above 175°C, softens, blackens and apparently polymerizes above 195°C, and finally melts in the range 310-335°C. NMR analysis: 1H -NMR (DMSO- d_6/D_2O , 300 MHz) δ (ppm) from TMS: m (6.723, 6.741, 6.752); m (7.511, 7.518, 7.533, 7.539, 7.547, 7.559, 7.577, 7.585); m
35 (7.663, 7.674, 7.686, 7.698, 7.707); d (7.818, 7.846). In the absence of D_2O , the compound also displays a broad singlet at 11.90 ppm. The isomeric components were analytically

resolved by thin-layer chromatography (silica gel plates, ethyl acetate solvent), giving two bands, R_f 0.82 and R_f 9.72. Mass spectrum for R_f 0.82: m/z (relative intensity): 174 (M^+ , 100), 160 (26.8), 144 (93.0), 117 (90.8), 97 (21.9), 89 (96.1), 71 (24.1). High resolution data for the M^+ peak: calculated for $C_9H_6N_2O_2$: 174.042928; found: 174.043200 (deviation = -0.3 ppm). For the component having R_f 0.72, M^+ , calculated for $C_9H_6N_2O_2$: 174.042928; Found: 174.043200 (deviation = -1.6 ppm). These data confirm that the 10 compounds are mono-nitroso isomers.

F. Iodo-Nitro Substituted Estrones

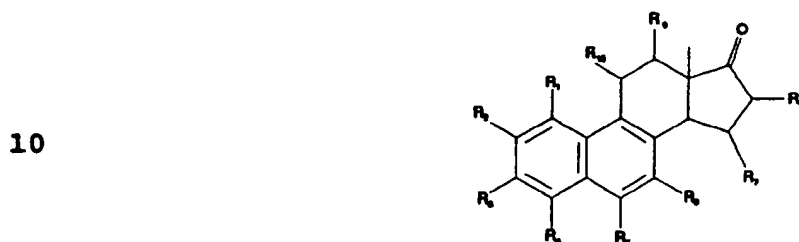
The method of total estrone synthesis of Bachman *et al.*, J. Amer. Chem. Soc. 64:974 (1942) is used for the synthesis of iodo-nitro substituted estrones. These products are compounds of the following formula:



wherein, R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten , substituents are hydrogen, and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro.

G. Iodo-Nitro Substituted Equilenins

The method of Bachman *et al.*, J. Amer. Chem. Soc. 61: 974 (1939) is used for the synthesis of iodo-nitro substituted equilenins. These products are compounds of the following formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo (C_1 - C_6) alkyl, (C_1 - C_6 alkoxy, (C_3 - C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the nine R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 substituents are hydrogen, and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents are nitroso or nitro.

H. 2-Iodo-5-Nitroso-Benzamide

The synthesis of 2-iodo-5-nitroso-benzamide can be performed similarly to the synthesis of 4-iodo-3-nitroso-benzamide, described above, using 2-iodo-5-nitro benzoic acid (Chemica Alta Ltd., Edmonton, Alberta, Canada) as the starting material.

I. 2-Iodo-5-Nitro-Benzamide

2-iodo-5-nitro-benzamide was synthesized by the method previously described for 4-iodo-5-nitro-benzamide using 2-iodo-5-nitro benzoic acid (Chemica Alta Ltd., Edmonton, Alberta, Canada) as the starting material.

¹H NMR Spectrum, in DMSO-d₆ δ (ppm) values relative to TMS: 7.742 (1H, singlet); 7.943 (1H, doublet of doublets, J=8.26 Hz, J=2.58 Hz); 8.034 (1H, singlet), 8.064 (1H, doublet, J=2.94 Hz); and 8.191 (1H, doublet, J=8.46 Hz).

5

Mass Spectrum:

Low resolution electron impact spectrum (m/z): 292 (M⁺), 276, 230, 202, 165, 127, 91, 75, 63. High resolution measurement of M⁺: Calculated for C₇H₅IN₂O₃: 291.934494; 10 found: 291.934149 (deviation= 1.2 ppm).

Melting Point: melting point 230-233°C.

J. 4-Iodo-3-Aminobenzamide

15 INO₂BA (730 mg, 2.50 mmol) was dissolved in warm EtOH (25 mL) to which was then added H₂O (25 mL) at 24°C followed by 1 M sodium dithionite (50 mL, freshly prepared in 0.05 M sodium bicarbonate). After 15 min. the product was extracted into ethyl acetate which was then removed by rotary 20 evaporation, and the residue crystallized from hot H₂O (40 mL). Yield: 139 mg (21%). This amino compound does not form a hydrochloride.

Microanalysis: Calculated for C₇H₅IN₂O: C, 32.08; 25 H, 2.69; I, 48.43; N, 10.69. Found: C, 31.57; H, 2.37; I, 48.70; N, 10.02.

¹H NMR Spectrum, in DMSO-d₆, δ (ppm): 5.30 (2H, multiplet, amino protons), 6.79 (1H, dd, J=8.2 Hz, J=1.9 Hz), 30 7.17 (1H, s), 7.23 (1H, d, J=1.9 Hz), 7.60 (1H, d, J=8.0 Hz), 7.75 (1H, s).

UV absorption spectrum, λ max (ε): 322, 230 nm and shoulders at 252, 220 nm.

35

Melting point: melting point 175-180°C (decomp.).

K. 6-Amino-1,2-benzopyrone

The synthesis of 6-amino-1,2-benzopyrone is reported in Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991).

5 L. 5-Iodo-6-Amino-1,2-Benzopyrone

The synthesis of 5-iodo-6-amino-1,2-Benzopyrone is reported in Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991).

10 M. Further Examples

Compounds of formulae IV-IX are synthesized in a manner similar to Examples I.F. and I.G., above.

15 In a preferred mode compounds I-IX are synthesized with an iodo group adjacent to a nitro or nitroso group. In a more preferred mode, compounds I-IX are synthesized with an iodo group adjacent to a nitro group.

20 In an even more preferred mode compounds of formulae I and II are selected from the group consisting of 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide and 5-iodo-6-nitro-1,2-benzopyrone.

25

30

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II. I₅₀ and LD₁₀₀ Values for Nitroso- and Nitro- Compounds in (A) Mammary Cancer Cells (monolayer cultures) and in (B) L-1210 Murine Leukemia Cells (suspension culture).

The effect of nitroso- and nitro- compounds in mammary cancer cells and in murine leukemia cells is shown in Table 1.

TABLE 1

(A)		(Nanomoles/cm x 10 ⁴ cells)					
Cell line:		MDA 468		MCF-7		BT 474	
		I 50	LD100	I 50	LD100	I 50	LD100
10	NOBA						
	INO ₂ BA	85	195	100	250	90	200
	4-iodo-3-NO ₂ -benzoic acid	180	n.d.	145	n.d.	160	n.d.
	INO ₂ BP	260	n.d.	250	n.d.	260	n.d.

15 Seeding was at 2x10⁴ or 0.8 x 10⁴ cells/cm², drugs were added 6-18 hours after seeding, and drug exposure time was 24-48 hours.

(B)		L 1210 Cells (nanomoles/10 ⁵ cells)	
		I 50	LD100
20	NOBA	7.5	15
	NO ₂ BA	16	32
	4-Iodo-3-NO ₂ -benzoic acid	16	32
	NO ₂ BP	35	80

Cells were seeded at 5 x 10⁴ to 2 x 10⁵/ml; drugs were added at the time of seeding; drug exposure was for 18 hours.

25 III. Comparison of pADPRT Inactivation and pADPRT Zinc Content by NOBA and INOBA

NOBA and INOBA were assayed for their respective effects on pADPRT enzyme inactivation and pADPRT zinc content according to the methods described in Buki et al., FEBS Lett. 30 290:181-185 (1991) at pH 7.0. pADPRT inactivation was performed in 20-μl volumes of buffer containing 50 mM Hepes (pH 7.4), 100 mM KCl, 0.5 mM EDTA, pADPRT (0.4 μg/μl) and different concentrations of inactivators. Inactivators were serially diluted in the above buffer from a 20 mM stock solution in DMF. The DMF concentration (2.5%) was kept 35 constant in the inactivation mixes. After 2 hr at 25°C,

aliquots (3 μ l) were removed for assay of enzyme activity in a volume of assay mix (200 μ l) which the inactivators were dilute enough not to act as inhibitors. The assay mix consisted of 100 mM Tris (pH 7.7), 14 mM 2-mercaptoethanol, 5 0.1 mM [32 P]-NAD $^{+}$, 0.2 mg/ml coDNA and 0.1 mg/ml histones. The result of the pADPRT assays are shown in Figure 1. The incorporated 32 P, proportional to enzyme activity, is plotted as percent of the untreated sample. The effect of NOBA and INOBA on the zinc content of pADPRT is shown in Figure 2. In 10 these experiments 65 Zn $^{2+}$ -labeled pADPRT was used.

The Zn $^{2+}$ content of pADPRT (Figure 2A, upper panel) was determined as follows: two parallels of aliquots (4 μ l) were pipetted into 500 μ l of ice-cold washing solution (50 mM Tris, pH 7.7, 10 mM 2-mercaptoethanol and 0.5 mM EDTA) then 15 filtered onto GF/C disks presoaked in washing solution (cold), quickly washed 4 times with 1 ml of ice-cold washing solution, dried, and the radioactivity determined. The protein-bound 65 Zn $^{2+}$ is shown as percent of the untreated sample. Enzyme assays (Figure 2A, lower panel) were 20 performed as in Figure 1.

For the release of Zn $^{2+}$ from transblotted pADPRT, the enzyme (3 μ g per lane) was separated on SDS-PAGE, transblotted onto a nitrocellulose membrane, and renatured in the presence of 65 ZnCl $_2$. The Zn $^{2+}$ -loaded transblot was cut 25 into strips and incubated with 2 ml of 50 mM Hepes/NaOH (pH 7.0) and 0.5 mM EDTA containing different concentrations of either NOBA or INOBA as indicated, for 2 hr. at 25°C. After incubation, 1 ml of the supernatant was counted for radioactivity, and the strips were washed 3 times with 2 ml 30 of 50 mM Tris-HCL pH 7.4), 0.5 mM EDTA and 10 Mm 2-mercaptoethanol, and then dried and exposed to X-ray film. The intensity of the autoradiographic spots was quantitated by a scanner. The results are shown in Figure 2B.

IV. Effectiveness of 4-(halogen)-3-nitro-benzamide compounds as tested with human T-lymphoblastoid leukemia cell lines in culture.

	I_{50}	LD_{100}
5 4-iodo-3-nitro-benzamide	70	190
4-fluoro-3-nitro-benzamide	120	280
4-chloro-3-nitro-benzamide	130	300
4-bromo-3-nitro-benzamide	120	290

10 Cells (MT-2 cells, or, in case of 3-fluoro-4-nitro-benzamide, CEM-4 cells) were seeded at a density of 0.1×10^5 /ml; the drugs were added at the time of seeding. The cultures were incubated at 37°C in a 5% CO₂ atmosphere and the cells numbers were monitored by hemacytometer counting in 24
15 hr intervals. " I_{50} " values (left column) represent the concentrations estimated to decrease cell viability to 50% of control; " LD_{100} " values (right) are concentrations that achieved zero viability after 48 hours of incubation.

20 V. Comparison of Cytostatic and Cytocidal Activity of 4-Iodo-3-Nitrobenzamide and 2-Iodo-5-Nitrobenzamide

The cytostatic and cytocidal activity of 4-iodo-3-nitrobenzamide and 2-iodo-5-nitrobenzamide are shown below:

	Cytstatic Effect I_{50} (E ras cells)	Cytocidal Effect I_{50} (with 0.8 mM BSO)
25 4-iodo-3-nitrobenzamide	60 μ M	4 μ M
2-iodo-5-nitrobenzamide	12 μ M	2 μ M

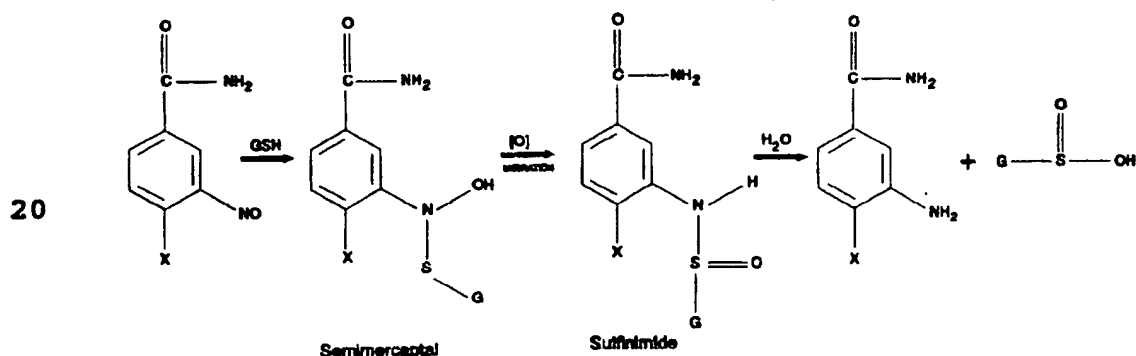
30 Each value is the average of three trials. Each trial deviated from the average by no more than twenty percent. The 2-iodo-5-nitrobenzamide isomer is the more effective of the two isomers tested at inducing cytostatic and cytocidal
35 effects. Another isomer of the iodo-nitro-benzamide series, 2-iodo-4-nitrobenzamide had no effect at the concentrations given above.

VI. Metabolic Conversion of INO₂BA to INH₂BA

It is known that GSH adds to aromatic nitroso compounds to form labile adducts (semimercaptals) one of whose reaction pathways is molecular rearrangement (oxygen atom migration) to yield sulfinamides that hydrolyse to the corresponding amines plus glutathione sulfinic acid (Equation 1). Eyer *et al.*, Chem. Biol. Interactions 24:227-235 (1979); Umemoto *et al.*, Chem. Biol. Interactions 68: 57-69 (1988); Ellis *et al.*, Chem. Biol. Interactions 82:151-163 (1992).

Equation 1

15



25 Where X=H or I;
 GSH is glutathione (γ - L - glutamyl - L - cysteinylglycine tripeptide);
 G — S — denotes the glutathion - S - yl group in the semimercaptal; and
 G — S(O) — denotes the glutathione sulfinyl group in the oxidized species

30

The cell metabolites of INO₂BA metabolism were identified. A suspension of 855-2 cells (6), 150 x 10⁶ cells in 30 ml of RPMI-1640 medium, was dosed with INO₂BA to a concentration of 120 μM, incubated at 37°C for 18 hr; and then freeze-dried, extracted 3 times with EtOH (10 ml each time) for 30 min and centrifuged. The extract was rotary

evaporated to dryness, redissolved in EtOH (3.0 ml), microfuged to remove small particles, and aliquots (200 μ l) injected into HPLC. In control experiments, instead of incubating the drug-dosed cell suspension it was immediately frozen, freeze-dried and extracted. HPLC analyses were performed essentially as reported earlier, except the pH of the buffers was 4.3. Buki *et al.*, FEBS Lett. **290**:181-185 (1991). Upon sample injection the gradient started from 100%A to 100%B in 30 min, to 50%B/50%C in 15 min, to 100%C in 3 min. Retention times of synthetic standards were: 4-iodo-3-hydroxylaminobenzamide 18.7 min; 4-iodo-3-aminobenzamide ("INH₂BA") 24.7 min; INO₂BA 34.5 min; INOBA 36.5 min and 6,6'-diiodo-3,3'-dicarbamoylazoxybenzene 46.9 min.

A slow but apparently steady reduction of the nitro to the amino end-product occurred at a rate of $2.2 \pm 0.2\%$ per 18 hrs of cell incubation at 37°C. Apart from as yet unidentified trace metabolites the only identified metabolite was INH₂BA (*see* Equation 1, where X=I). The removal of GSH by BSO augments the rate of accumulation of INOBA from INO₂BA by decreasing the rate of removal by GSH, as deduced from the pathway shown in Equation 1.

VII. Cytocidal Effect of NOBA and INO₂BA in Combination With Ascorbate

Because of the *in vitro* reactivity of the nitroso group of INOBA with both ascorbate and GSH, the apoptosis-inducing effect of NOBA with the action of the prodrug INO₂BA in the presence of ascorbate, added simultaneously with both drugs was compared. As shown in Table 2, 160 μ M ascorbate, which by itself had no effect on AA-2 leukemia cells, completely protected against the apoptosis-inducing action of NOBA, but did not prevent apoptosis by INO₂BA after 18 hr of drug exposure. These results are readily explained by the rapid chemical reduction of the nitroso group in NOBA to the hydroxylamine, and the rapid cellular oxidation of ascorbate to its oxidized form. Since the generation of INOBA from

its prodrug is slow and steady, it would be expected that the apoptosis-inducing capacity of the prodrug is sustained for the entire duration of the incubation (18 hr) whereas the rapid oxidative removal of the ascorbate takes place in 5 minutes, thus its protective effect is of short duration only. However these results predict that a chemotherapeutic efficiency of the prodrug would be much greater in species that do not synthesize ascorbate.

10

TABLE 2

Cytocidal Effect of NOBA and INO₂BA in combination with Ascorbate

		Cell number (x10 ⁶ /ml)	
		Day 1	Day 2
15	Control	0.51	0.90
	Ascorbate (160 μ M)	0.48	0.90
20	NOBA (32 μ M)	0.29	0.37 (50% dead)
	NOBA + Ascorbate	0.57	0.91
	INO ₂ BA (16 μ M)	0.25	0.45 (40% dead)
25	INO ₂ BA + Ascorbate	0.27	0.50 (40% dead)

AA-2 cells were seeded at a density of 0.2 x 10⁶ cells/ml. Drugs and/or sodium ascorbate were added at the time of seeding, and cells were counted by hemacytometer after incubation (37°C, 5% CO₂) for 18 hr (Day 1) and for 42 hr (Day 2). Viability was evaluated by Trypan Blue exclusion. Assays were performed in triplicates and deviation from averages did not exceed $\pm 15\%$.

30

35

VIII. Synergistic Cytotoxic Effects of 4-Iodo-3-Nitrobenzoic Acid, INO₂BA and NOBA, Respectively, in Combination with BSO

A time course of the cell killings by 4-iodo-3-nitrobenzoic acid, INO₂BA and NOBA, each alone and in combination with BSO was obtained. Molt-4 cells were seeded at a density of 0.05×10^6 cells/ml in the recommended growth medium in the absence or presence of drugs as indicated in Figure 3. Drugs were added to the wells immediately after seeding from 10-40 mM stock solution in DMSO. BSO was added from a 100 mM aqueous stock solution. Cells were incubated at 37°C in a 5% CO₂ atmosphere and proliferation was monitored in 14 hours by cell counting in a hemacytometer. The results of the experiment are shown in Figure 3.

As is apparent from Figure 3, 4-iodo-3-nitrobenzoic acid, INO₂BA and NOBA had no significant effect on the cell proliferation of either Molt-4 cells or L-1210 cells over a 2-3 day period. BSO alone also had no significant effect on cell proliferation during this period. BSO in combination with each of the drugs showed rapid synergistic cytotoxic effects in both Molt-4 and L-1210 cells as followed for 2-3 days.

Inasmuch as we find that some malignant cell types contain pADPRT which is not auto-poly-ADP-ribosylated in the intact cell, Cole et al., Biochem Biophys. Res. Commun 180:504 (1991), a structural analog of INOBA not containing the carboxamide group, 4-iodo-3-nitrosobenzoic acid (via its nitro precursor), would not be expected to bind to pADPRT, because only the carboxamide group effectively binds at the nicotinamide site. Althaus and Richter, ADP-Ribosylation of Proteins: Enzymology and Biological Significance, Springer-Verlag, Berlin (1987). The majority of tumor cells respond to the benzamide-containing prodrug much more readily than to the carboxylic acid. That reduction of the prodrug to its nitroso species is essential for induction of apoptosis is evident from the large activating effect of BSO, which by diminishing cellular GSH decreases the rate at which GSH

chemically reduces, and thus inactivates, the nitroso molecules generated.

IX. Synergistic Effect of INO₂BA in Combination with BSO on Cytotoxicity of Various Cancer Cell Lines

The role of cellular GSH was determined by the simultaneous addition of INO₂BA and BSO (an inhibitor of GSH biosynthesis, Meister, Pharmacology and Therapeutics 51:155-194 (1991), which at the concentrations applied diminished cellular GSH levels below 5-10% of controls.) The results are tabulated below in Table 3.

TABLE 3

Synergistic effect of DL-buthionine sulfoximine (BSO) on cytotoxicity of INO₂BA in various cancer cell lines

No.	Cell Line	I ₅₀ (μM)		LD ₁₀₀ (μM)	
		INO ₂ BA	INO ₂ BA + BSO	INO ₂ BA	INO ₂ BA + BSO
1	MDA-468	80	15	180	40
2	MCF-7	100	16	200	30
3	MCF-7-B-ADR	90	16	220	45
4	Du 145	120	20	250	50
5	HT 144	100	15	200	35
6	Hep-G2	120	20	200	45
7	PC-12	60	16	120	50
8	L 1210	128	20	256	64
9	CEM-4	80	15	196	32
10	HL 60	70	8	125	16
11	Molt-4	70	15	196	32
12	U 937	65	8	150	16

The concentration of BSO was 1 mM for all cell lines except HT 144, where the concentration was 0.25 mM. Cells were seeded in the recommended growth medium at a density of 1.5 x 10⁴ cell/cm² into 2 cm² wells. Drugs were added simultaneously 4 hours after seeding, and incubation was continued for 48 hours at 37°C in a 5% CO₂ atmosphere. For assessment of drug effects cells were detached by trypsinization and counted in a hemacytometer.

Cell lines Nos. 1-4 were obtained from Dr. C. Benz, University of California, San Francisco. MDA 468 and MCF-7 human mammary cancer lines, MCF-7-B-ADR drug-resistant mammary cancer line, Du 145 drug-resistant human prostate cancer line, HT 144 human melanoma cell line, Hep-G2 human hepatocellular carcinoma line, and PC-12 rat pheochromocytoma line were obtained from the University of California Cell Culture Facility. Each value in Table 3 is an average of two parallels, each of which did not deviate from the average by 10 more than $\pm 15\%$.

As summarized in Table 3 seven types of human cancer cells cultured as monolayers and five types of leukemia cells grown as suspension cultures responded to BSO by a dramatic increase in the apoptosis-inducing potency of INO_2BA . Notable 15 are the MCF-7-B-ADR adriamycin-resistant human mammary cancer cell line and the Du-145 drug-resistant prostatic cancer cell line. That reduction of the C-nitro precursor to the active C-nitroso compound is essential for induction of apoptosis is evident from the large synergistic effect of BSO, which by 20 diminishing cellular levels of GSH decreases the rate at which GSH chemically reduces, and thus inactivates, the active C-nitroso compounds generated in vivo.

X. Non-Toxicity of INO_2BA and BSO

25 Twelve hamsters of both sexes were injected intraperitoneally with 200 mg/kg INO_2BA daily for 14 days without toxic effects.

The non-toxicity of BSO has been reported in the literature. See, e.g., Liebman *et al.*, Cancer Res. 53:2066-30 2070 (1993); Yao *et al.*, Cancer Res. 53:3662-3666 (1993); Terradez *et al.*, Biochem. J. 292:477-483 (1993); and Polhuijs *et al.*, Biochem. J. 285:401-404 (1992).

XI. Zinc loss from Treatment of HIV-1 Virions with NOBA

35 Experiments were performed to determine if NOBA is capable of ejecting zinc from intact virions. HIV-1 (MN

strain) was produced, purified and concentrated as described in Bess *et al.*, J. Virol. 66:840-847 (1992). The concentrated virus was diluted to 60 times that of culture fluid in TNE buffer (0.01 M Tris-HCl, 0.1 M NaCl, 1 mM EDTA, pH 7.2) and incubated with 3000 or 6000 μ M NOBA at 37°C. The virus was then pelleted and washed with TNE buffer to remove weakly bound zinc. The quantity of zinc in the resulting viral pellets was determined as described in Bess *et al.*, J. Virol. 66:840-847 (1992). No significant loss of viral proteins in the pellet was detected by p24 and gp120 competition radioimmunoassays and coumassie-stained sodium dodecyl sulfate polyacrylamide gel electrophoresis.

The data in Table 4 demonstrate that treatment of concentrated suspensions of HIV-1 (60x with respect to culture solution) with NOBA results in losses of 50-83% of the viral zinc and complete loss of infectivity. Since edge x-ray absorption fine structure spectroscopy has shown that the majority of the zinc in intact retroviruses is coordinated by the CCHC ligands, Summers *et al.*, Protein Science 1:563-574 (1992) and Chuang *et al.*, Proc. Natl. Acad. Sci. 89: (1992), the ejection of zinc from virions by NOBA is directly attributable to a destabilization of the nucleocapsid CCHC zinc fingers.

TABLE 4

25

NOBA (μ M)*	Incubation time (hr)	Zinc, Control Sample (μ g/ml)	Zinc, NOBA-Treated Sample (μ g/ml)	Zinc Loss (%)
3,000	2	0.21	0.11	52
6,000	4	0.24	0.04	83

* concentrations correspond to molar NOBA:zinc finger ratios of ca. 350:1 (3,000 μ M) and 700:1 (6,000 μ M)

30

XII. In Vitro Anti-Retroviral Testing of 6-NOBP and NOBA

Both 6-NOBP and NOBA were tested on viral growth of HIV-1 (LAV strain) in phytohemagglutinin-stimulated human peripheral lymphocytes (PBL). An HIV-1 stock having an infectivity titer of 100,000 TCID₅₀ was incubated for 30 min. at 22°C with either of the drugs. Control HIV samples,

containing no drugs, were incubated in the same manner. Following successive serial dilutions of 10-fold that resulted in HIV-1 titers as given in the lower abscissa of Figure 7 (A,B), viral growth was initiated by adding the HIV-1 dilutions to PBL and allowing an incubation period of 9 days. At the end of this incubation cultures were assayed for productive infection by an immunoassay for HIV-1 antigens and by reverse transcriptase as described in McDougal *et al.*, J. Immunol. Methods 76:171-183 (1985). Virus titers were expressed as percentile values (percent infected cultures) compared to the controls, containing HIV dilutions which were not preincubated with the C-nitroso drugs. In a separate series of experiments, HIV-1 dilutions and C-nitroso drugs (see upper abscissa) were not preincubated but were added simultaneously to lymphocytes in exactly the same concentrations as described previously (i.e., following preincubation) and viral growth monitored 9 days later. As illustrated in Figure 8, the inhibition of HIV-1 propagation was profound when C-nitroso drugs were preincubated with HIV-1.

XIII. Anti-Viral Activity of C-Nitro- and C-Nitroso-Containing Compounds

TABLE 5

HIV Activity in Human Lymphocyte of pADPRT Ligands Calculated Infections Titer of Virus After Treatment

Treatment	Dose (μ M)	TCID ₅₀	Decrease	Inhibition
No Drug	No Drug	177,828	-	-
NO ₂ BP	500	85,114	0.32	52.14
NOBP	100	102	3.24	99.94
NOBA	100	11	4.20	>99.99

6-Nitro-1,2-benzopyrone ("NO₂BP"), an *in vivo* pro-drug of NOBP inhibited HIV-1 activity in human lymphocytes 52% by reduction to the active nitroso form *in vivo*. Since the nitro compound is more stable and soluble than the active

nitroso compound, which in the solid form is a poorly soluble dimeric species, supplying the nitro pro-drug provides a constant in vivo supply of the active nitroso compound for inhibiting viral growth and reproduction.

5

XIV. Synergistic Anti-Viral Effects of BSO and INO₂BA

CEM x174 cells (3×10^5 /ml) were infected with 3.75 TCID₅₀/ml (50% tissue culture infectious dosage per ml of cell suspension) stock solution of SIVmac239 or virus isolates
10 from SIVmac239-infected rhesus macaques on day 0. Uniform infection of cells was ensured by the infection of one cell population only per virus preparation. Three days post-infection, the cells were distributed (500 μ l/well) into individual wells of a 24-well plate. Fresh medium containing
15 INO₂BA concentrations ranging from 0 to 300 μ M and/or BSO concentrations ranging from 0 to 0.25 mM were added to establish final 1-ml reaction volumes, and the cultures were further incubated for 4 days. The wells were then replenished with fresh CEM x 174 cells (3×10^5 /ml), INO₂BA
20 and/or BSO were re-added at the same concentrations as in the original incubates, and incubation was continued for an additional 3 days. This procedure intends to mimic conditions of in vivo chemotherapy. Cell viability and virus titers in the supernatant were determined on day 10 (3 days
25 after the addition of new cells) by the tetrazolium (MTT) method of Hansen et al., J. Immunol. Methods 119:208-210 (1982) and by the SIV p27 and reverse transcriptase (RT) assays described by Chuang et al., FEBS Lett. 326:140-144 (1993), respectively.

30 The viability of infected and uninfected cell cultures was determined by the method of Hansen et al., J. Immunol. Methods 119:208-210 (1982). Briefly, MTT was dissolved in sterile phosphate buffered saline (PBS) at a concentration of 5 mg/ml and 20 μ l of MTT solution was added into each
35 microtiter well containing 100 μ l of cell culture. After 2 hr incubation at 37°C, 100 μ l of solubilizing solution was added. Following overnight incubation at 37°C, optical

absorbance were measured at 570 nm using a microtiter plate reader (Molecular Devices Corp., Sunnyvale, CA).

To evaluate the activity of SIV reverse transcriptase, 10 μ l of infected cell supernatant was added to a reaction mixture containing 50 mM Tris-HCl (pH 8.0), 5 mM $MgCl_2$, 10 mM dithiothreitol, 20 mM KCl, and 1% Triton X-100 in a total volume of 50 μ l. Poly(rA):oligo (dT)₁₂₋₁₈ was present at 100 μ g/ml and 2.4 μ M [³H]TTP was used. The reaction mixtures were incubated at 37°C for 1 hr and the TCA-precipitable radioactivity was filtered onto nitrocellulose filters which were then washed, dried and counted.

SIVmac p27 core antigen levels were determined by an enzyme immunoassay provided by Coulter Corp. (Hialeah, FL). The assay was performed according to the manufacturer's specifications.

The results of these experiments are shown in Figures 4 and 5. Just like the inhibition of syncytia formation by the drug, apparent cell viability (tested by the MTT assay) was protected by INO_2BA , indirectly indicating a depression of viral replication. Four SIV strains behaved essentially in a similar manner, and INO_2BA increased cell viability proportional to decreasing viral replication, with BSO supporting this effect. See Figures 4A and 4B.

A more direct anti-SIV activity is illustrated in Figures 5A, 5B and 5C, where a decrease to almost complete inhibition of SIV p27 formation is apparent as a function of INO_2BA concentration. By itself, BSO had no effect on p27 formation (Fig. 5D). With a combination of BSO and varying concentrations of INO_2BA (10, 50 and 250 μ M), in various SIV strains, p27 formation was markedly depressed, an effect that roughly correlated with the inhibition of syncytia formation. As an independent assay of viral replication, reverse transcriptase activity of SIV-infected CEM x174 cells was greatly depressed by INO_2BA (Fig. 6).

Just as in tumor cells where a slow reduction of the prodrug INO_2BA to the bioactive $INOBA$ produces cell killing, an anti-SIV effect of the prodrug is apparent in the CEM x174

cell line. This cell line, requiring no stimulation by phytohemagglutinin for viral growth, is a fusion product of human B cell line 721.174 and T cell line CEM. Slater et al., Immunogenics 21:235-246 (1985). As is the case with many established immortal cell lines it does not exhibit a normal phenotype since it is immortal. If the assumption that undifferentiated cells have an impaired NADPH oxidase system, Trudel et al., Biochem J. 278:611-619 (1964), is valid for CEM x 174 cells, this defect also predisposes these cells to reduce INO_2BA to the active nitroso species, contrary to the behavior of normal cells containing intact NADPH oxidase. However, present results indicate that replication of HIV in intact human macrophages, which presumably have no defective NADPH oxidase, is readily inhibited by the prodrug INO_2BA . Kun et al., Abstract 422A, Tenth International Conference on AIDS, Yokohama, Japan (1994). This poses the question whether retroviral infection by itself is capable of interfering with NADPH oxidase. If so, this condition may promote the selectivity of INO_2BA towards HIV or SIV infected cells in vivo. The highly effective synergistic action of BSO with INO_2BA strongly supports the contention that the nitroso species (INOBA) exerts its direct action on retroviral CCHC zinc fingers, as reported earlier, Rice et al., Nature 361:473-475 (1993), because depression of cellular GSH by BSO produces a higher, and thus more effective, concentration of the nitroso drug in the cell.

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XV. The Effect of INO₂BA on SIVmac Replication in CEM x174 Cells Pre-Infected with Virus Before Treatment with INO₂BA

The effect of INO₂BA on SIVmac replication in CEM x174
5 cells that were pre-infected with virus for one day prior to
treatment with INO₂BA are shown in Figure 6. The cell culture
and SIV reverse transcription assay were performed as
described in Example XIV.

This experiment clearly indicates that INO₂BA is
10 effective even when incubated with cells that have been
infected with virus prior to INO₂BA addition, whereas NOBA
requires pre-incubation with cells prior to viral infection.

XVI. Synergistic Effect of INO₂BA and BSO on AZT-Resistant SIV Propagation
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The synergistic effect of INO₂BA in combination with BSO on AZT-resistant SIV propagation is shown in Table 6.

TABLE 6

5 The Synergistic effect of INO₂BA and BSO on AZT-resistant SIV propagation

	Virus	INO ₂ BA (μM)	BSO (mM)	Syncytia
10	SIV 23686	0	0	+++
	(AZT-resistant)	25	0	++
		50	0	++
		100	0	++
		200	0	+
		300	0	-
		0	0.01	+++
		25	0.01	+
		50	0.01	+
		100	0.01	-
15		200	0.01	-
		300	0.01	-
		0	0.01	-
		25	0.01	-
		50	0.01	-
		100	0.01	-
		200	0.01	-
		300	0.01	-
		0	0.01	-
		25	0.01	-
20	SIV 22803	0	0	+++
	(AZT-resistant)	25	0	++
		50	0	++
		100	0	++
		200	0	-
		300	0	-
		0	0.05	++++
		25	0.05	-
		50	0.05	-
		100	0.05	-
25		200	0.05	-
		300	0.05	-
		0	0.05	-
		25	0.05	-

30 CEM x 174 (3 x 10⁵/ml) were infected with equal doses of SIVmac239 or virus isolates from SIVmac239-infected rhesus macaques (MMU 23686, 22803). Three days post-infection, INO₂BA concentrations ranging from 0 to 300 μM with or without BSO were added, and incubated for an additional 3 days. Cell cultures were then examined for syncytia formation. The number of syncytia in cell cultures was counted in arbitration fields under 40X magnification and scored as follows: over 30 (++++), 20-30 (+++), 10-20 (++), 1-9 (+), and 0 (-).

The infection of CEM x 174 cells with SIVmac virus results in syncytia formation and eventual cell death by lysis. Chuang *et al.*, Biochem. Biophys. Res. Commun. 195:1165-1173 (1993). In 4 days an AZT-resistant SIV strain (SIV-23686) induced between 20-30 syncytia per visual field.

However increasing concentrations of INO_2BA (from 25 to 300 μM) effectively abolished cell fusion above 200 μM drug concentration. A second AZT-resistant strain (SIV-22803) produced over 30 syncytia and required higher than 100 μM INO_2BA to abrogate cell fusion. When BSO was added simultaneously with the drug, the inhibitory effect on syncytia formation was greatly enhanced, and as little as 0.01 mM BSO (in SIV-23686) lowered the inhibitory concentration of INO_2BA to 50 μM , whereas 0.05 mM BSO decreased the syncytia-blocking concentration INO_2BA to 25 μM (in SIV-22803). Differences between strains may be related to intracellular GSH, which is probably affected differently by the two viral strains.

XVII. Synergistic Cytotoxic Effect of IABP in Combination with INO_2BA in L 1210 Cells

The synergistic effect of IABP in combination with INO_2BA in L 1210 cells is shown in Figure 10. As can be seen in the Figure, 160 μM , 320 μM and 480 μM IABP decreased L 1210 cell proliferation by approximately 55%, 80% and 93%, respectively, over a three day growth period. 64 μM INO_2BA decreased L 1210 cell proliferation by approximately 80% over a three day growth period. 64 μM in combination with 160 μM , 320 μM and 480 μM IABP, respectively, decreased L 1210 proliferation by about 90%, 98% and 97%, respectively, demonstrating the synergistic cytotoxic effect of IABP in combination with INO_2BA .

XVIII. Inhibitory Effect of IABP on SIVmac239 Replication

The effect of IABP on the infection cycle of SIV was determined. CEM x174 cells ($3 \times 10^5/\text{ml}$) were infected with SIVmac239 on day 0 and incubated for 3 days (37°C, 5% CO_2) before treatment with IABP (0-0.5 mM). The cultures were incubated for an additional 4 days before being replenished with fresh CEM x174 cells ($3 \times 10^5/\text{ml}$) and IABP. On day 10 the levels of free virus in the supernatants of the cultures

were assessed using the SIV p27 core antigen assay (Coulter Copr.) The absorbance was measured at 450 nm.

The results are shown in Figure 8. IABP has no toxic effect on cells, Cole et al., Biochem. Biophys. Res. Commun. 5 180:504-514 (1991), and depresses p27 levels proportional to its concentration.

XIX. Synergistic Anti-Retroviral Effect of IABP in Combination with INO₂BA

10 The synergistic effect of IABP in combination with INO₂BA on SIVmac239 replication was demonstrated. CEM x174 cells (1 x 10⁶/ml) were infected with SIVmac239 on day 0 and incubated for 3 days (37°C, 5% CO₂) before treatment with INO₂BA (0-300 μM) (rear columns) or with INO₂BA and IABP (0.3 mM) (front
15 columns). The cultures were incubated for an additional 4 days before being replenished with fresh CEM x174 cells (3 x 10⁵/ml) and drug(s). On day 10 the levels of free virus in the supernatants of the cultures were assessed using the SIV p27 core antigen assay (Coulter Copr.), with absorbance
20 measured at 450 nm. The results are shown in Figure 9A.

The synergistic effect of IABP in combination with INO₂BA on the viability of SIVmac239-infected CEM x174 cells was also demonstrated. The procedure was the same as that described in the preceding paragraph. The cell viability of
25 the cultures was measured by the MTT assay on day 10 of the experiment. The results are shown in Figure 9B. The effect of INO₂BA alone is shown in the front columns, and the drug combination in the rear columns.

A more than additive anti-SIV action of 0.3mM IABP and
30 varying concentrations of INO₂BA, as assayed by the inhibition of p27 formation in SIV-infected CEM x174 cells, was observed. This drug combination also supported cell growth more than additively in the presence of SIV.

35 XX. Non-Toxicity of IABP

The non-toxicity of IABP is demonstrated in Cole et al., Biochem. Biophys. Res. Commun. 180:504-514 (1991).

EQUIVALENTS

5 The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. Indeed, various modifications of the above-described makes for carrying out the invention which are obvious to those skilled in the field of molecular biology or
10 related fields are intended to be within the scope of the following claims.

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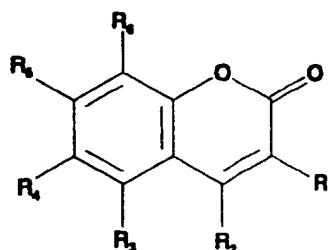
CLAIMS

What is claimed is:

1. A synergistic composition comprising one or a plurality
5 of pADPRT CCHC-oxidizing ligands in combination with one or a plurality of agents selected from the group consisting of: agents that decrease cellular levels of glutathione and non-covalent pADPRT-inhibitory ligands.
- 10 2. The composition of claim 1 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of:

a compound having the formula:

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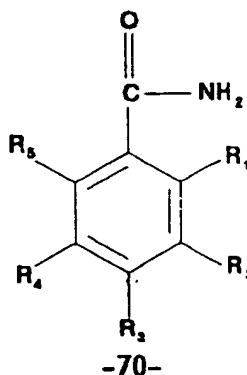
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- wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
25 salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro;

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a compound having the formula:

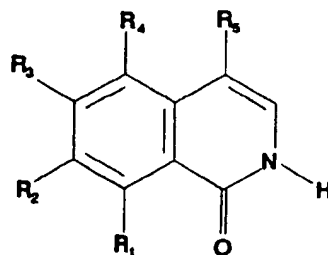
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wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one
5 another, selected from the group consisting of hydrogen,
hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4
and R_5 substituents are hydrogen and wherein at least one of
10 the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or
nitro;

a compound having the formula:

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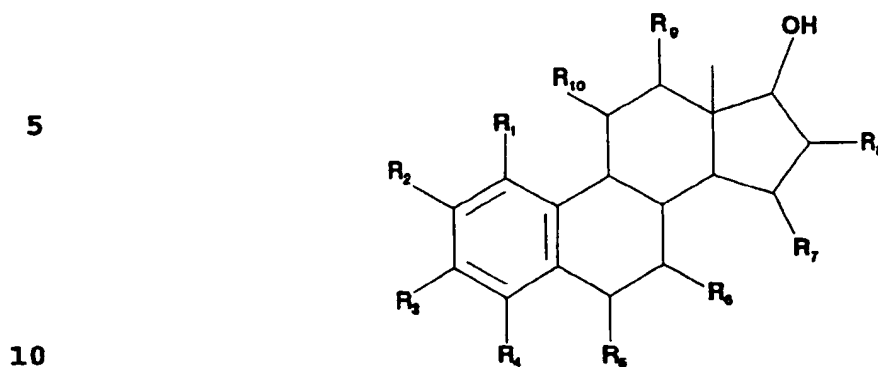
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wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one
another, selected from the group consisting of hydrogen,
hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
25 salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4
and R_5 substituents are hydrogen and wherein at least one of
the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or
nitro;

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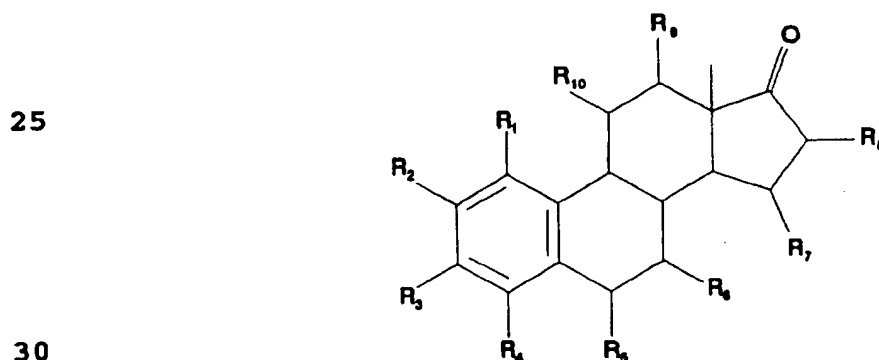
a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro;

20

a compound having the formula:

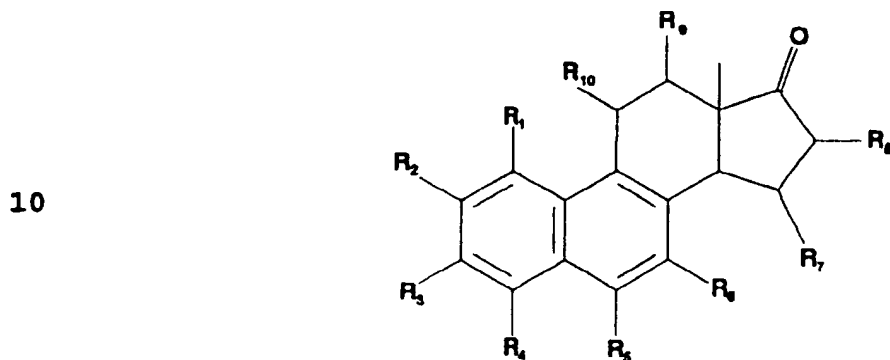


wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10}

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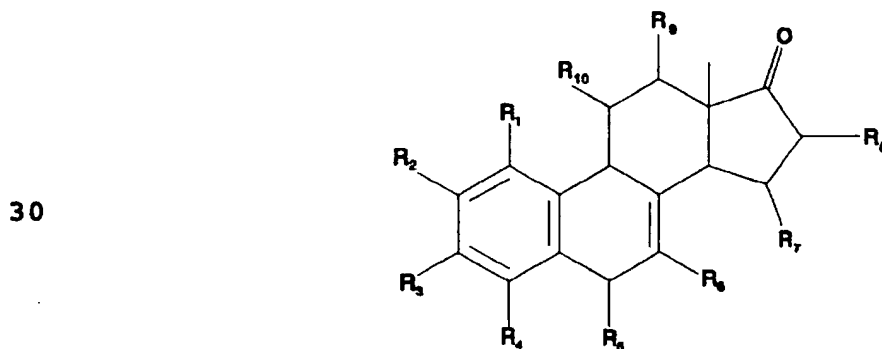
substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso or nitro;

5 a compound having the formula:



15 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least
20 three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso or nitro;

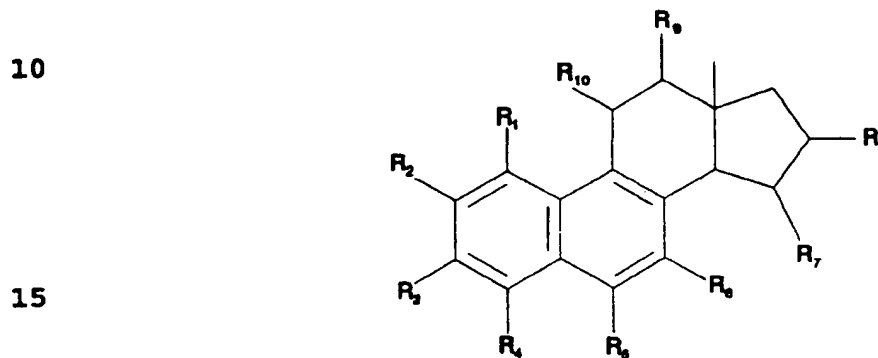
25 a compound having the formula:



35 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6)

alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents are hydrogen and wherein at least one of the ten
 5 R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents is nitroso or nitro;

a compound having the formula:

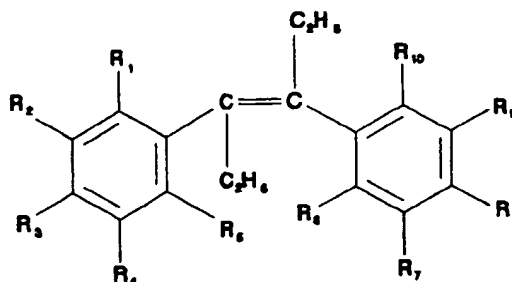


wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ are, independent of one another, selected from the group
 20 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆) alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents are hydrogen and wherein at least one of the ten
 25 R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents is nitroso or nitro; and

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a compound having the formula:



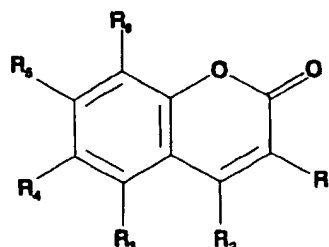
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wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and
 15 pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro.

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3. The composition of claim 2 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of:

a compound having the formula:



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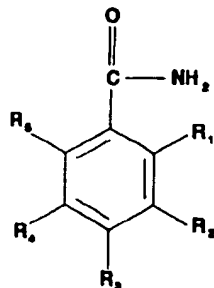
wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
 35 salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen and wherein at least

one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro; and

a compound having the formula:

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wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or nitro.

4. The composition of claim 3 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of 3-nitroso-benzamide, 4-iodo-3-nitroso-benzamide, 2-iodo-5-nitroso-benzamide, 3-nitro-benzamide, 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide, 6-nitroso-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, 6-nitro-1,2-benzopyrone, and 5-iodo-6-nitro-1,2-benzopyrone.

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5. The composition of claim 4 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide and 5-iodo-6-nitro-1,2-benzopyrone.

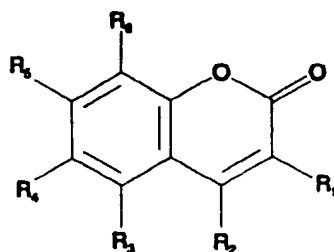
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6. The composition of claim 1 wherein said glutathione decreasing agents are selected from the group consisting of diethylmaleate and BSO.

5 7. The composition of claim 1, wherein said non-covalent pADPRT-inhibitory ligands are selected from the group consisting of:

a compound having the formula

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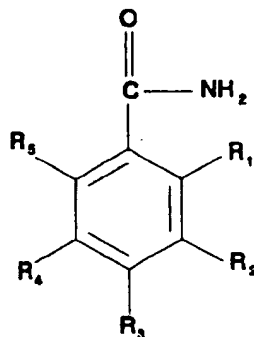
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wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, amino, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen; and

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a compound having the formula:

25



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wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, amino, iodo, alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts

35

thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen.

8. The composition of claim 7 wherein said non-covalent
5 pADPRT-inhibitory ligands are selected from the group consisting of 6-amino-1,2-benzopyrone and 5-iodo-6-amino-1,2-benzopyrone.

9. A pharmaceutical formulation comprising a composition
10 according to claim 1 and a pharmaceutical excipient.

10. A method of inactivating a retrovirus comprising the step of administering to said retrovirus a composition according to claim 1.

15

11. The method of claim 10 wherein said retrovirus is selected from the group consisting of HIV and SIV.

12. The method of claim 10 wherein said retrovirus is an
20 AZT-resistant retrovirus.

13. A method of inducing apoptosis in a tumor cell comprising the step of administering to a cell culture or mammalian host having said tumor cell a composition according
25 to claim 1.

14. A method for the treatment of cancer comprising the step of administering to a mammal an effective amount of a composition according to claim 1.

30

15. A method for the treatment of retroviral infections comprising the step of administering to a mammal an effective amount of a composition according to claim 1.

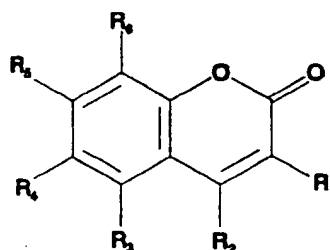
35 16. A synergistic composition comprising one or a plurality of pADPRT CCHC-oxidizing ligands in combination with one or a

plurality of agents that decrease cellular levels of glutathione.

17. The composition of claim 16 wherein said one or
5 plurality of pADPRT CCHC-oxidizing ligands are selected from the group consisting of:

a compound having the formula:

10



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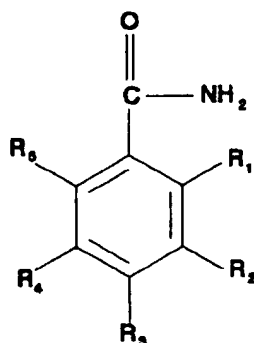
wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
20 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro;

25

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a compound having the formula:

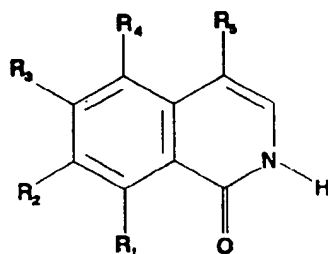


10

wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro;

20

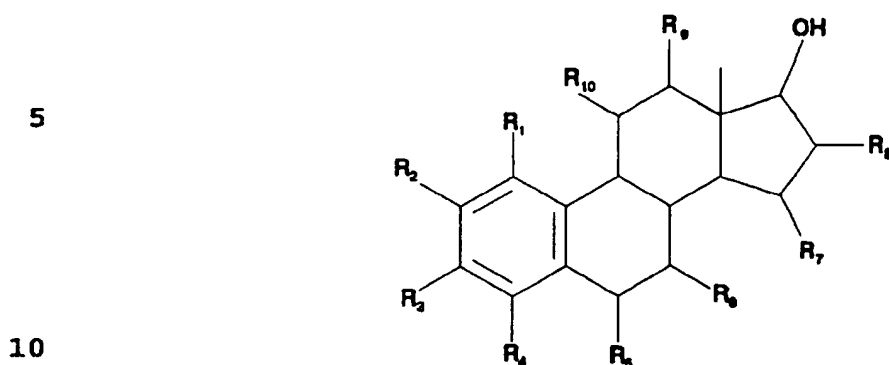
a compound having the formula:



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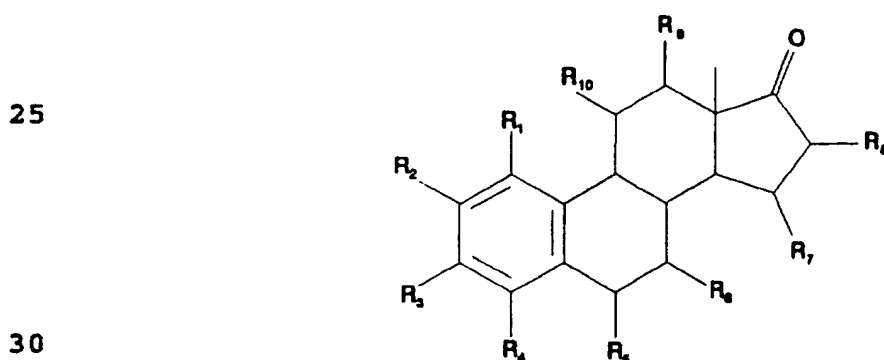
wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or nitro;

a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro;

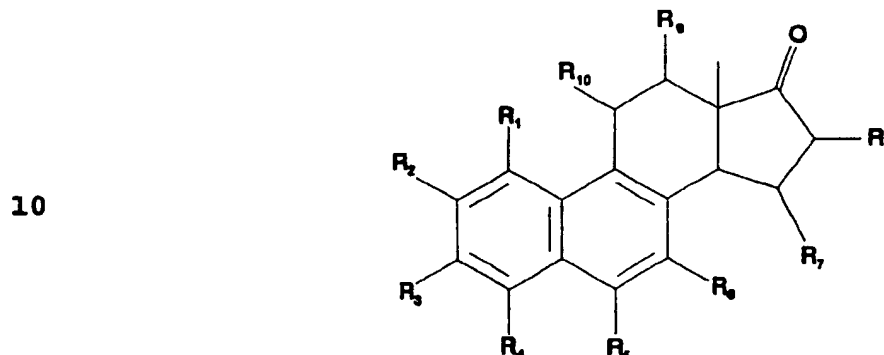
a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10}

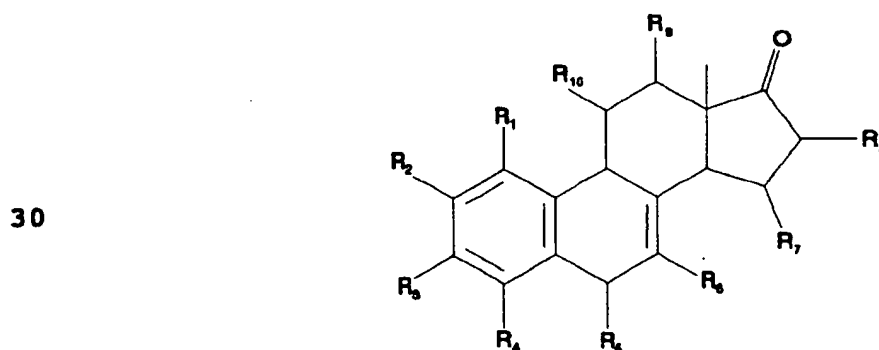
substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso or nitro;

5 a compound having the formula:



15 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least
20 three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso or nitro;

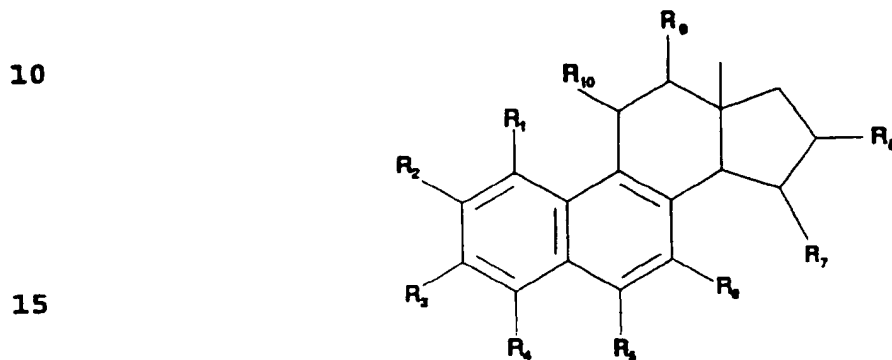
25 a compound having the formula:



35 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6)

alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents are hydrogen and wherein at least one of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso or nitro;

a compound having the formula:

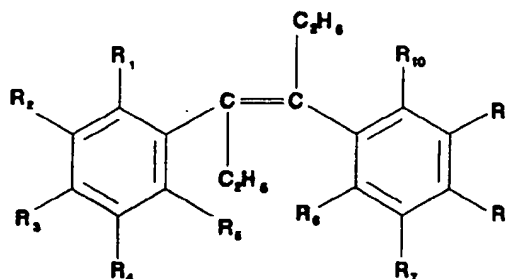


wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆) alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents are hydrogen and wherein at least one of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉ and R₁₀ substituents is nitroso or nitro; and

30

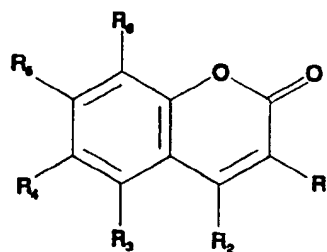
35

a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro.

18. The composition of claim 17 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of:
a compound having the formula:



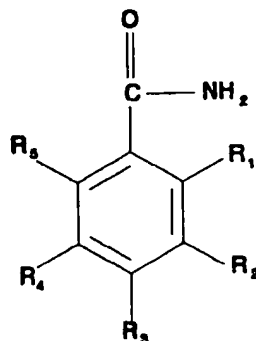
wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen and wherein at least

one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro; and

a compound having the formula:

5

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wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4 and R_5 substituents are hydrogen and wherein at least one of the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or nitro.

19. The composition of claim 18 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of 3-nitroso-benzamide, 4-iodo-3-nitroso-benzamide, 2-iodo-5-nitroso-benzamide, 3-nitro-benzamide, 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide, 6-nitroso-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, 6-nitro-1,2-benzopyrone, and 5-iodo-6-nitro-1,2-benzopyrone.

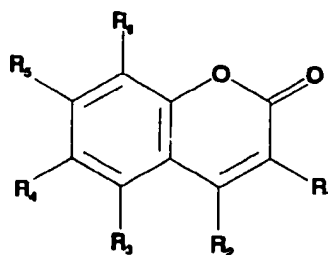
20. The composition of claim 19 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide and 5-iodo-6-nitro-1,2-benzopyrone.

35

21. The composition of claim 20 wherein said glutathione decreasing agents are selected from the group consisting of diethylmaleate and BSO.
- 5 22. A pharmaceutical formulation comprising a composition according to claim 16.
23. A method of inactivating a retrovirus comprising the step of administering to said retrovirus a composition
10 according to claim 16.
24. The method of claim 23 wherein said retrovirus is selected from the group consisting of HIV and SIV.
- 15 25. The method of claim 23 wherein said retrovirus is an AZT-resistant retrovirus.
26. A method of inducing apoptosis in a tumor cell comprising the step of administering to a cell culture or
20 mammalian host having said tumor cell an effective amount of a composition according to claim 16.
27. A method for the treatment of retroviral infections comprising the step of administering to a mammal an effective
25 amount of a composition according to claim 16.
28. A method of treating cancer comprising the step of administering to a mammal an effective amount of a composition according to claim 16.
30
29. A synergistic composition comprising one or a plurality of pADPRT CCHC-oxidizing ligands in combination with one or a plurality of non-covalent pADPRT-inhibitory ligands.
- 35 30. The composition of claim 29 wherein said one or plurality of pADPRT CCHC-oxidizing ligands are selected from the group consisting of:

a compound having the formula:

5

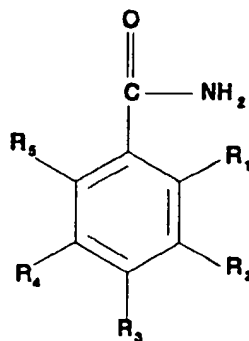


10 wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 ,
 15 R_4 , R_5 and R_6 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro;

20

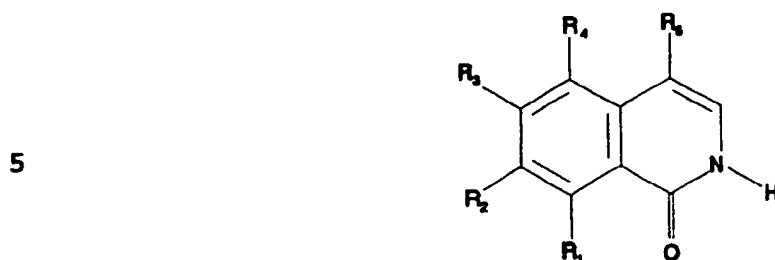
a compound having the formula:

25



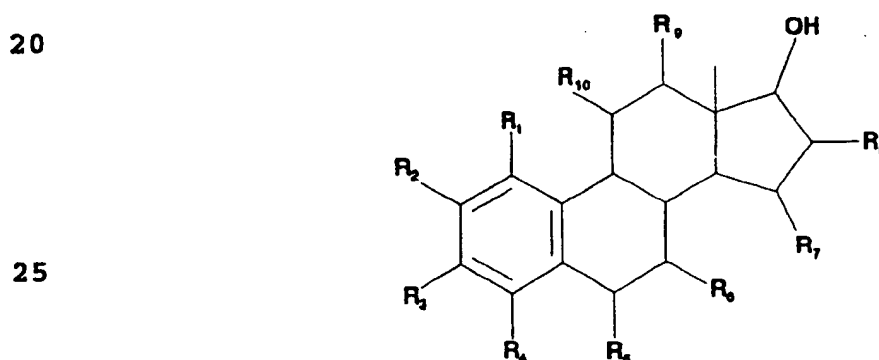
30 wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4
 35 and R_5 substituents are hydrogen and wherein at least one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso or nitro;

a compound having the formula:



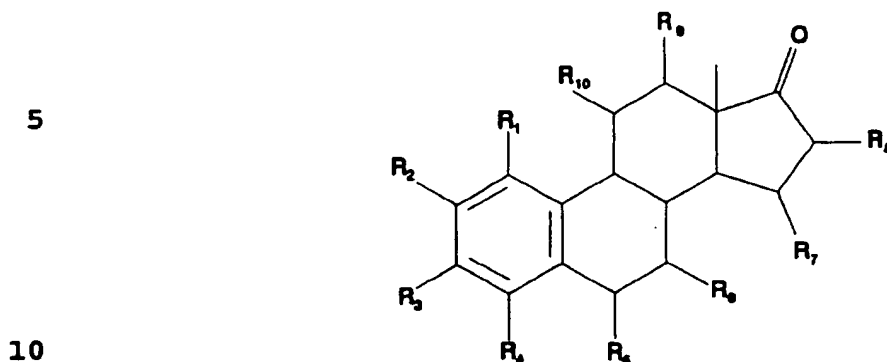
wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one
 10 another, selected from the group consisting of hydrogen,
 hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
 salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4
 and R_5 substituents are hydrogen and wherein at least one of
 15 the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or
 nitro;

a compound having the formula:



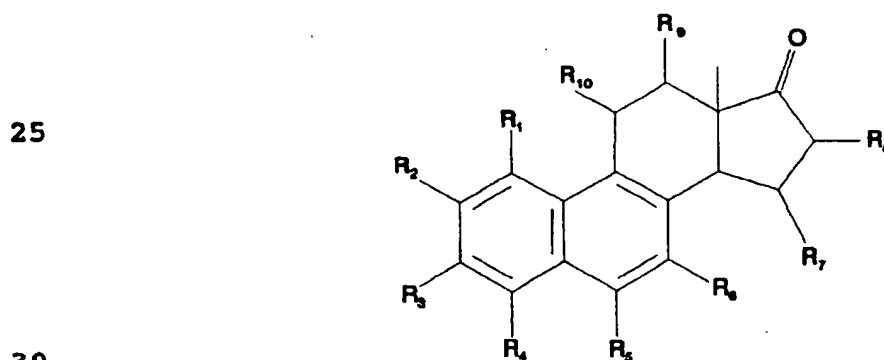
wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are,
 30 independent of one another, selected from the group
 consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6)
 alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and
 pharmaceutically acceptable salts thereof, wherein at least
 three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10}
 35 substituents are hydrogen and wherein at least one of the ten
 R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso
 or nitro;

a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and
 15 pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} substituents is nitroso
 20 or nitro;

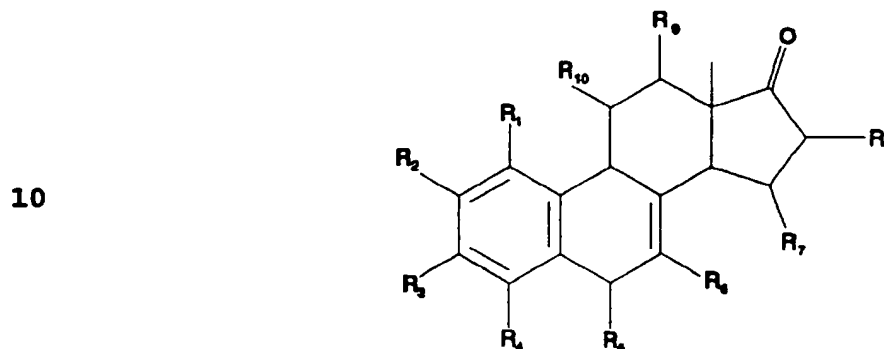
a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and
 35 pharmaceutically acceptable salts thereof, wherein at least three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 , and R_{10}

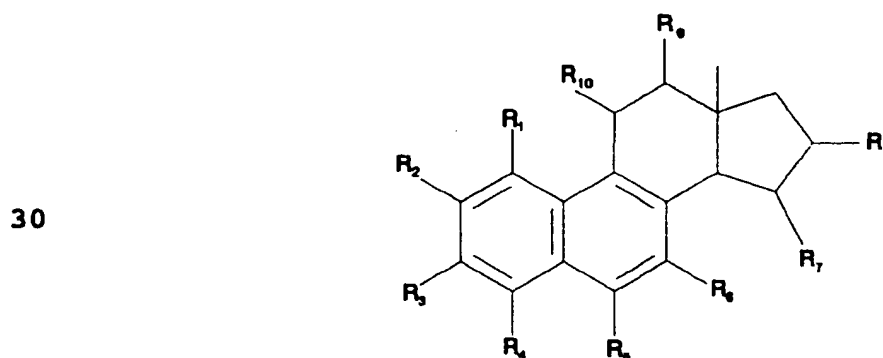
substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro;

5 a compound having the formula:



15 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least
20 three of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents are hydrogen and wherein at least one of the ten R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} substituents is nitroso or nitro;

25 a compound having the formula:

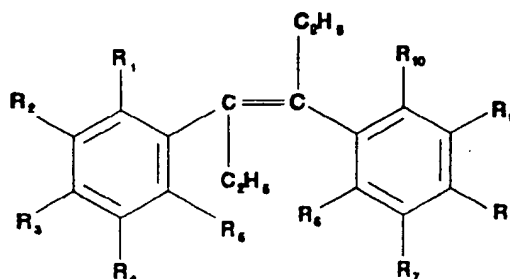


35 wherein R_1 , R_2 , R_3 , R_4 , R_5 , R_6 , R_7 , R_8 , R_9 and R_{10} are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C_1-C_6)

alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents are hydrogen and wherein at least one of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents is nitroso or nitro; and

a compound having the formula:

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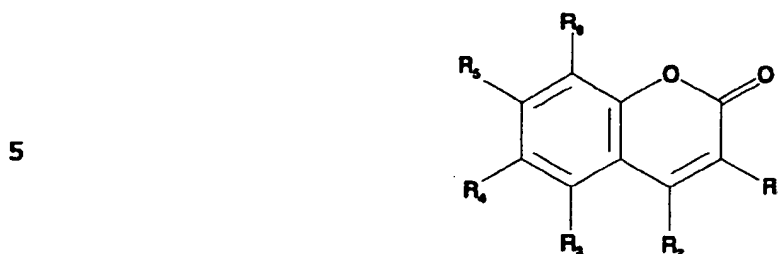
wherein R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ are, independent of one another, selected from the group consisting of hydrogen, hydroxy, nitroso, nitro, iodo, (C₁-C₆) alkyl, (C₁-C₆) alkoxy, (C₃-C₇) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents are hydrogen and wherein at least one of the ten R₁, R₂, R₃, R₄, R₅, R₆, R₇, R₈, R₉, and R₁₀ substituents is nitroso or nitro.

31. The composition of claim 30 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of:

30

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a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one
 10 another, selected from the group consisting of hydrogen,
 hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
 salts thereof, wherein at least three of the six R_1 , R_2 , R_3 ,
 R_4 , R_5 and R_6 substituents are hydrogen and wherein at least
 15 one of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents is nitroso
 or nitro; and

a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 and R_5 are, independent of one
 another, selected from the group consisting of hydrogen,
 hydroxy, nitroso, nitro, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy,
 30 (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable
 salts thereof, wherein at least two of the five R_1 , R_2 , R_3 , R_4
 and R_5 substituents are hydrogen and wherein at least one of
 the five R_1 , R_2 , R_3 , R_4 , and R_5 substituents is nitroso or
 nitro.

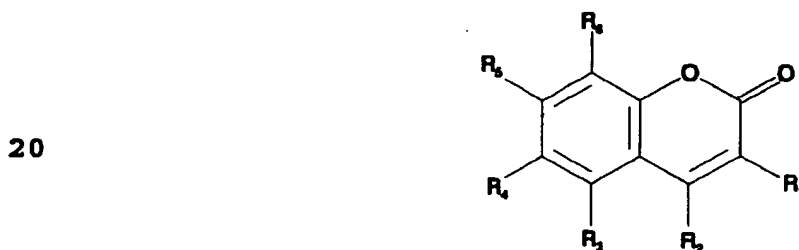
35 32. The composition of claim 31 wherein said pADPRT CCHC-
 oxidizing ligands are selected from the group consisting of

3-nitroso-benzamide, 4-iodo-3-nitroso-benzamide, 2-iodo-5-nitroso-benzamide, 3-nitro-benzamide, 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide, 6-nitroso-1,2-benzopyrone, 5-iodo-6-nitroso-1,2-benzopyrone, 6-nitro-1,2-5 benzopyrone, and 5-iodo-6-nitro-1,2-benzopyrone.

33. The composition of claim 32 wherein said pADPRT CCHC-oxidizing ligands are selected from the group consisting of 4-iodo-3-nitro-benzamide, 2-iodo-5-nitro-benzamide and 5-10 iodo-6-nitro-1,2-benzopyrone.

34. The composition of claim 29 wherein said non-covalent pADPRT-inhibitory ligands are selected from the group consisting of:

15 a compound having the formula

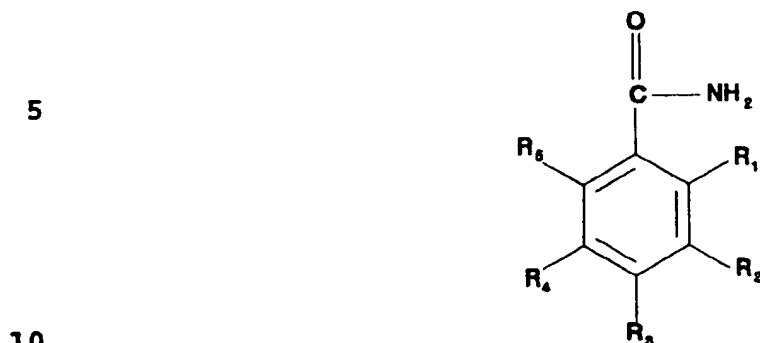


wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, 25 hydroxy, amino, iodo, (C_1-C_6) alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen; and

30

35

a compound having the formula:



wherein R_1 , R_2 , R_3 , R_4 , R_5 and R_6 are, independent of one another, selected from the group consisting of hydrogen, hydroxy, amino, iodo, alkyl, (C_1-C_6) alkoxy, (C_3-C_7) cycloalkyl, and phenyl and pharmaceutically acceptable salts thereof, wherein at least three of the six R_1 , R_2 , R_3 , R_4 , R_5 and R_6 substituents are hydrogen.

35. The composition of claim 34 wherein said non-covalent pADPRT-inhibitory ligands are selected from the group consisting of 6-amino-1,2-benzopyrone and 5-iodo-6-amino-1,2-benzopyrone.

36. A pharmaceutical formulation comprising a composition according to claim 29 and a pharmaceutical excipient.

37. A method of inactivating a retrovirus comprising the step of administering to said retrovirus a composition according to claim 29.

38. The method of claim 37 wherein said retrovirus is selected from the group consisting of HIV and SIV.

39. The method of claim 37 wherein said retrovirus is an AZT-resistant retrovirus.

40. A method of inducing apoptosis in a tumor cell comprising the step of administering to a cell culture or mammalian host having said tumor cell a composition according to claim 29.

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41. A method for the treatment of cancer comprising the step of administering to a mammal an effective amount of a composition according to claim 29.

10 42. A method of treating a retroviral infection comprising the step of administering to a mammal an effective amount of a composition according to claim 29.

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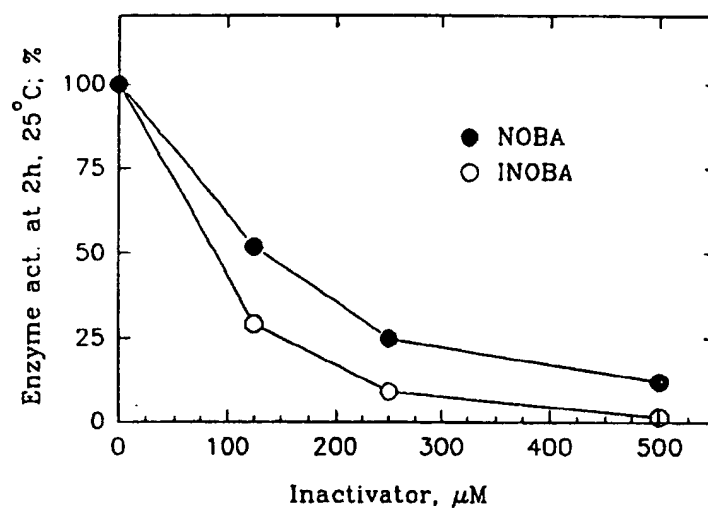


Figure 1

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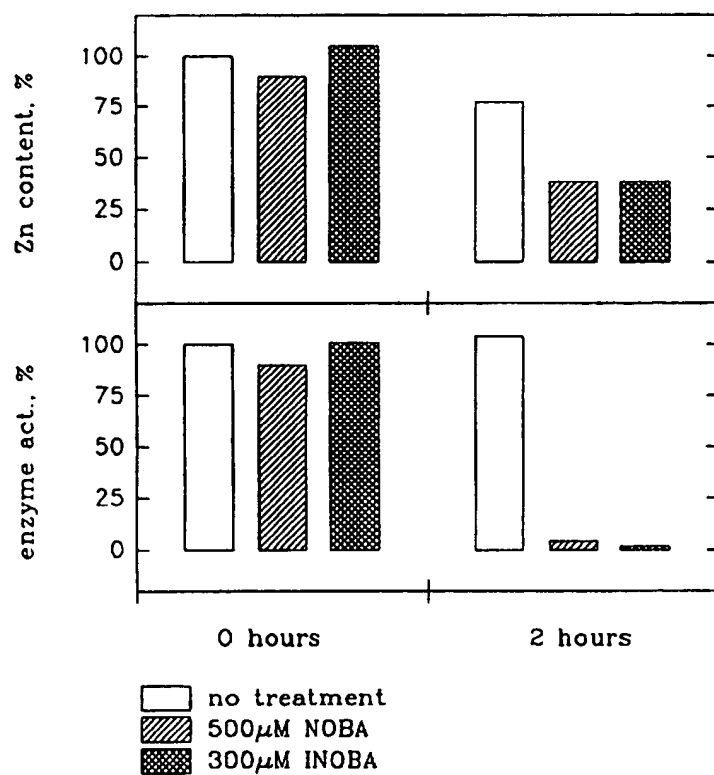


Figure 2A

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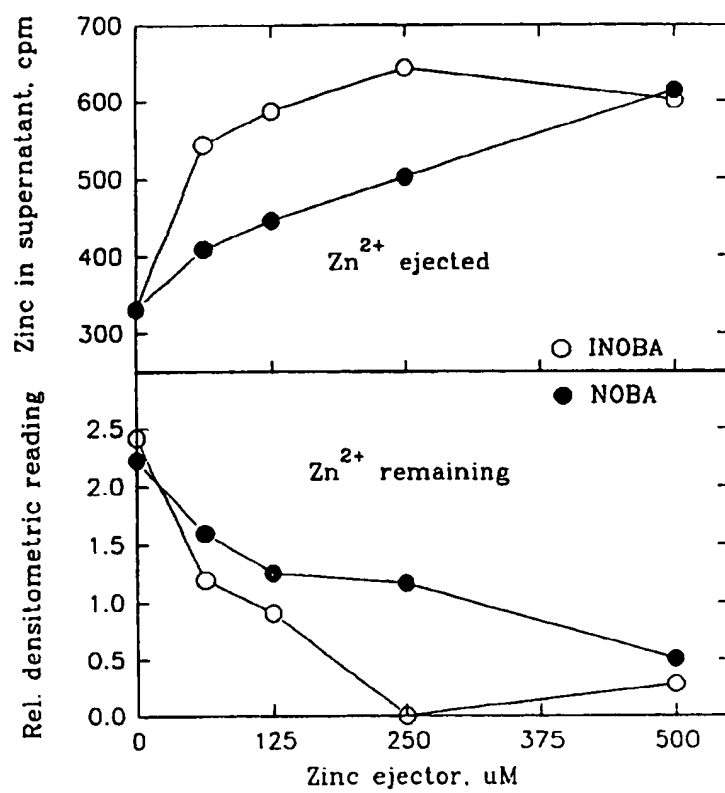


Figure 2B

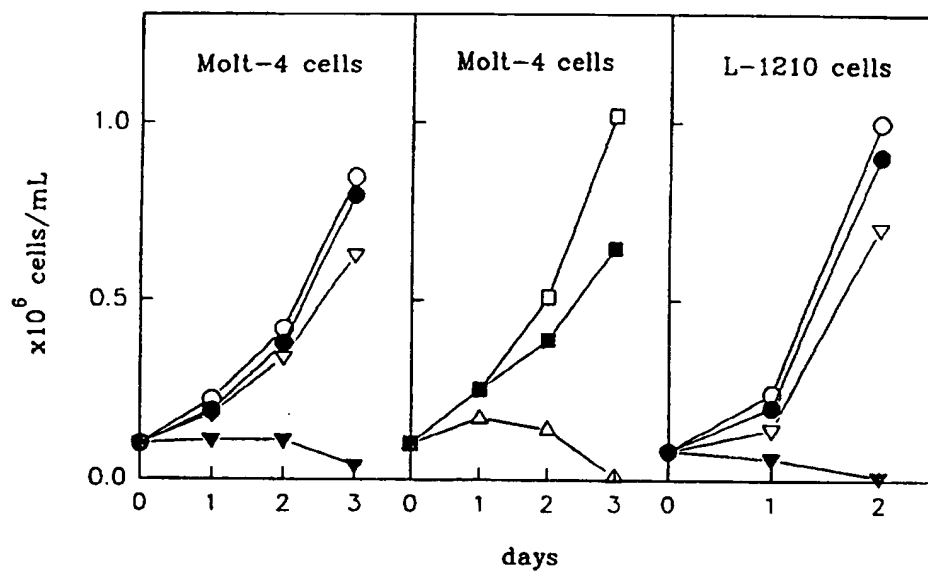


Figure 3

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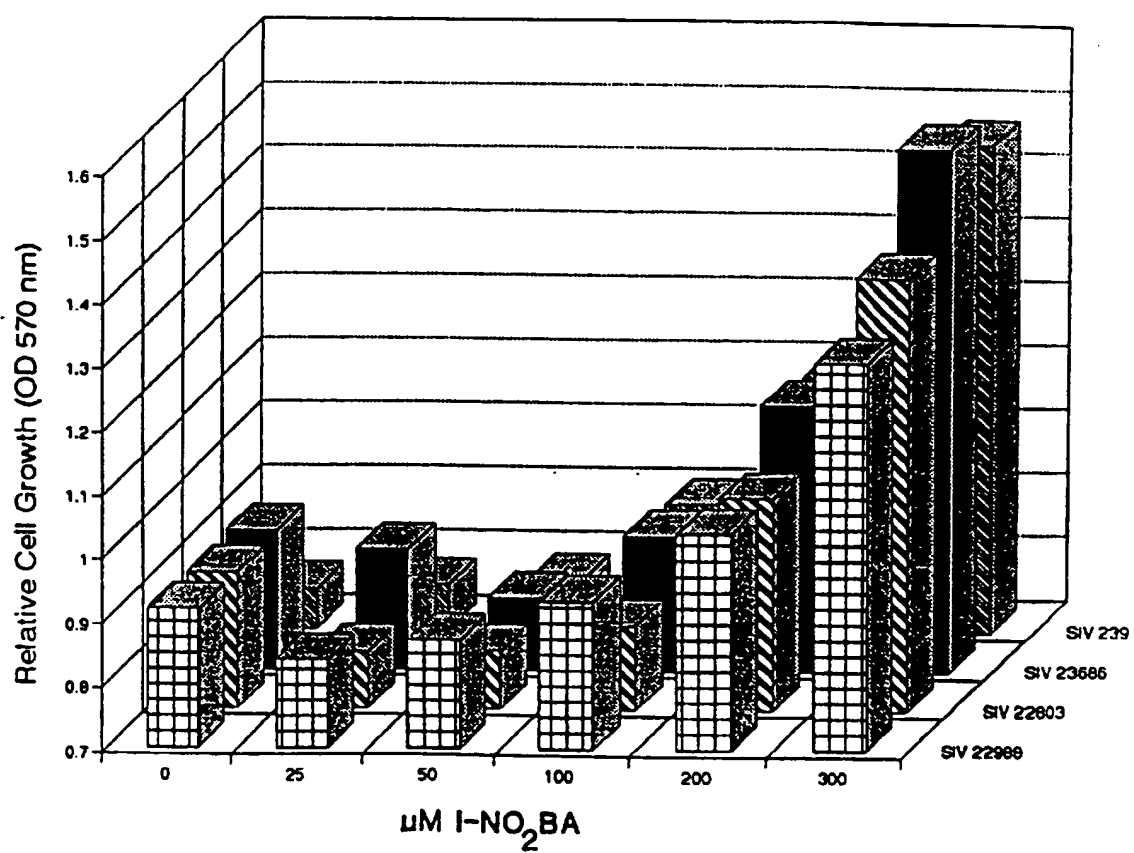


Figure 4A

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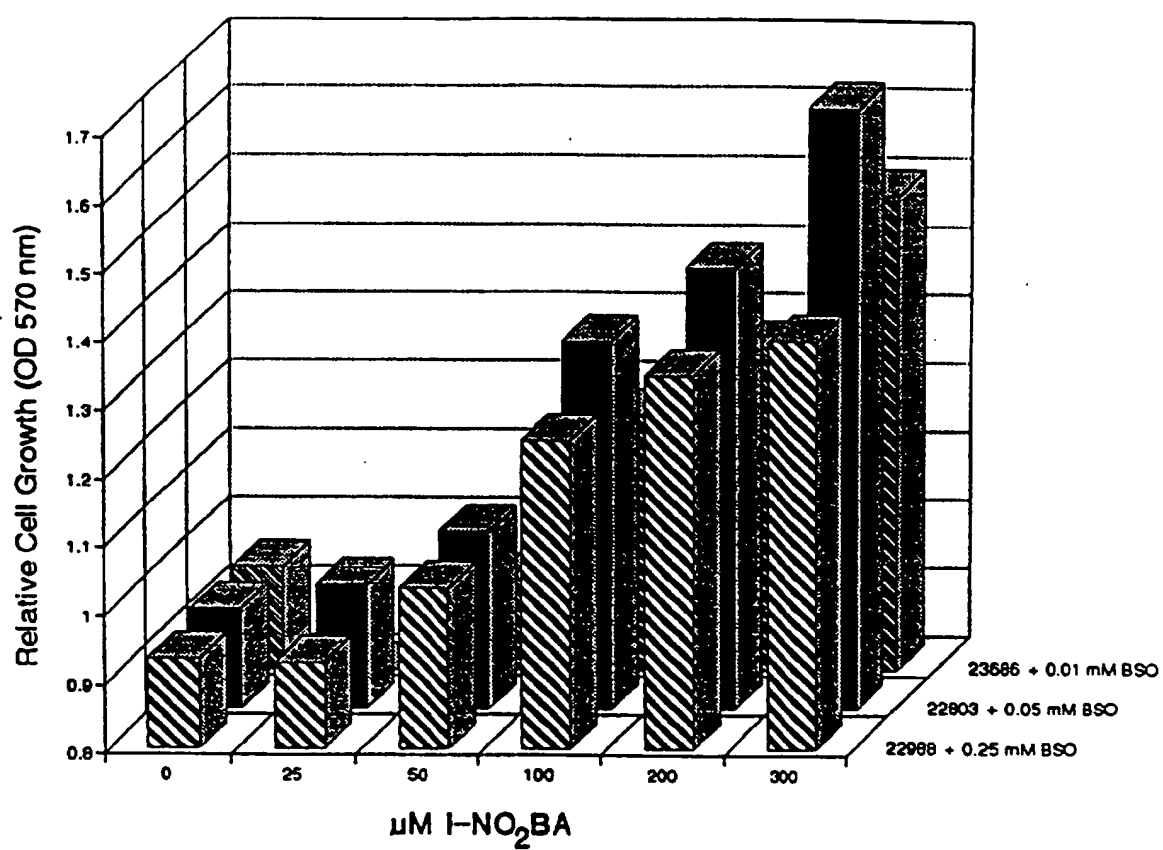


Figure 4B

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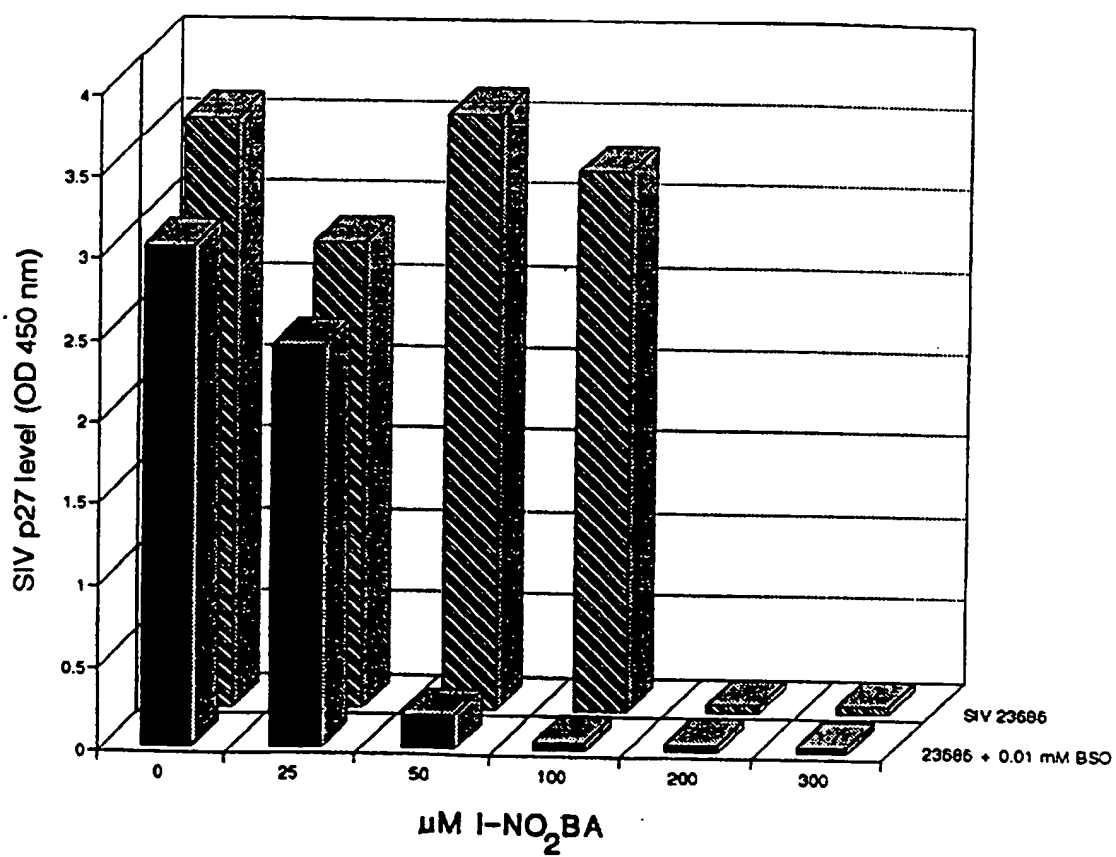


Figure 5A

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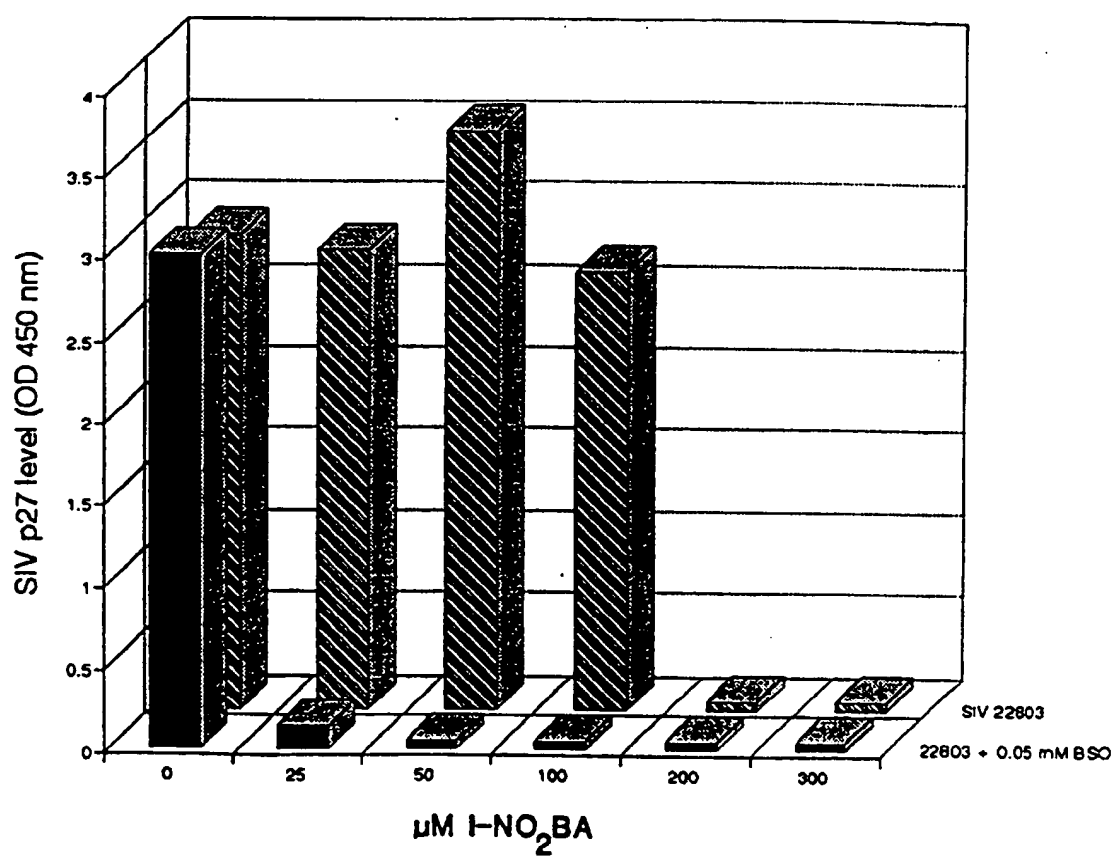


Figure 5B

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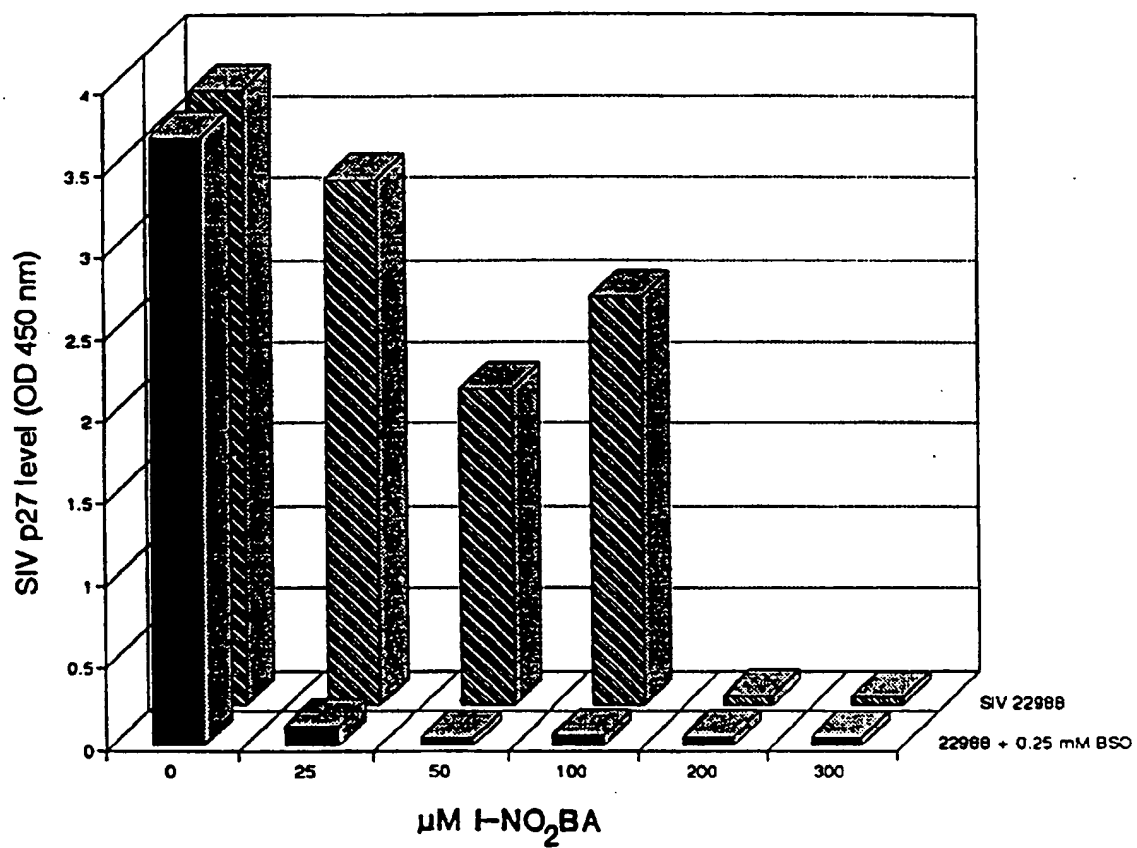


Figure 5C

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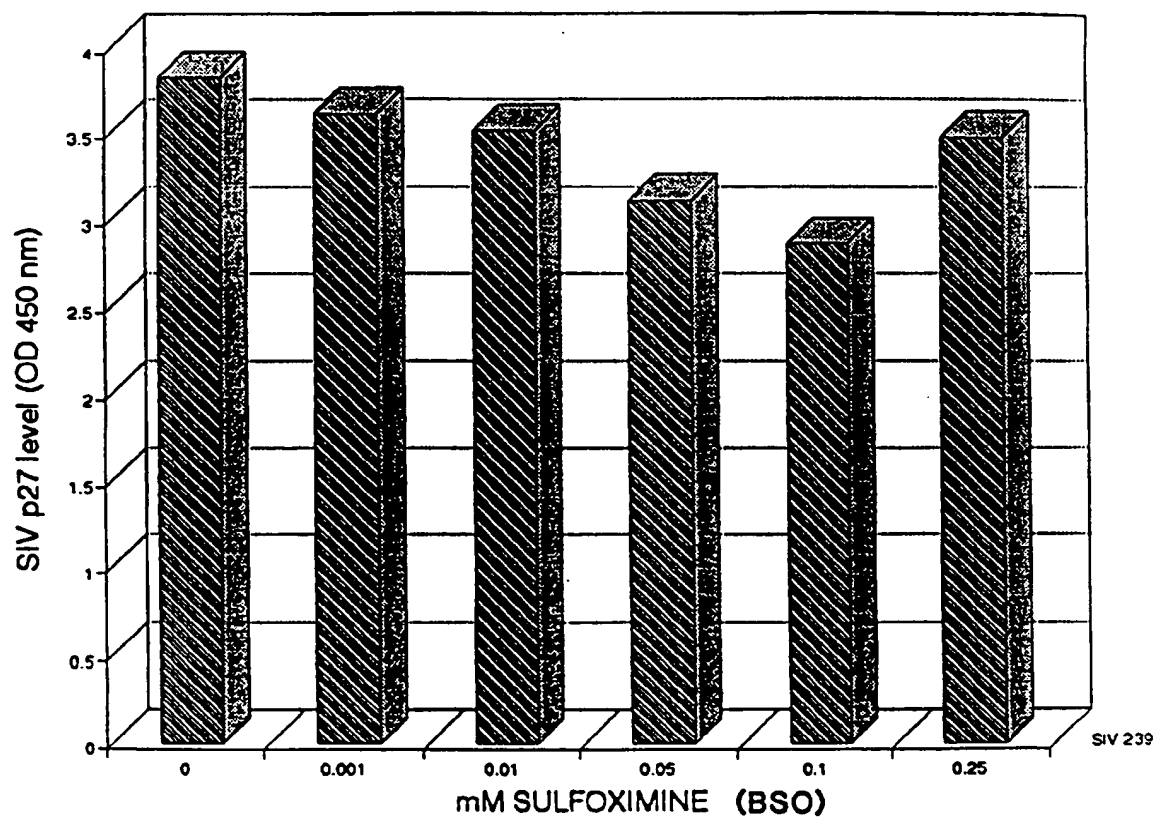


Figure 5D

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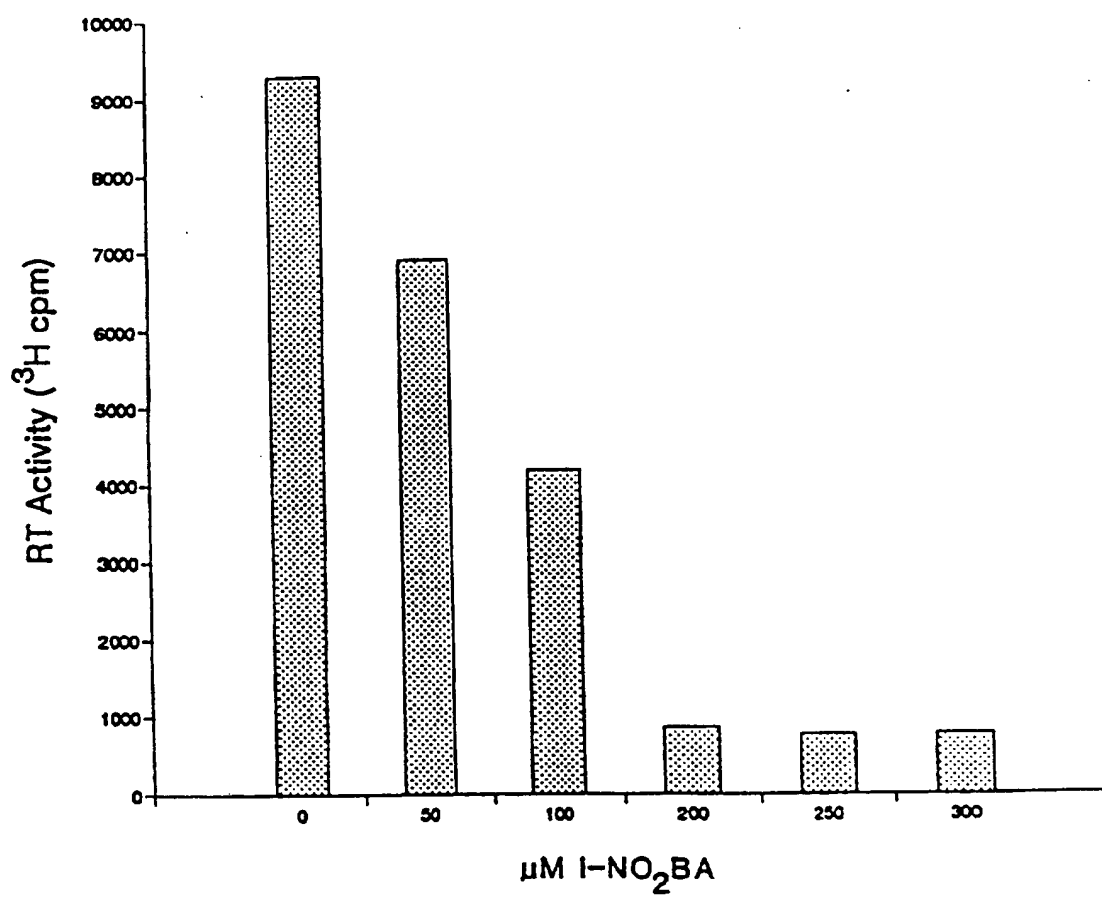


Figure 6

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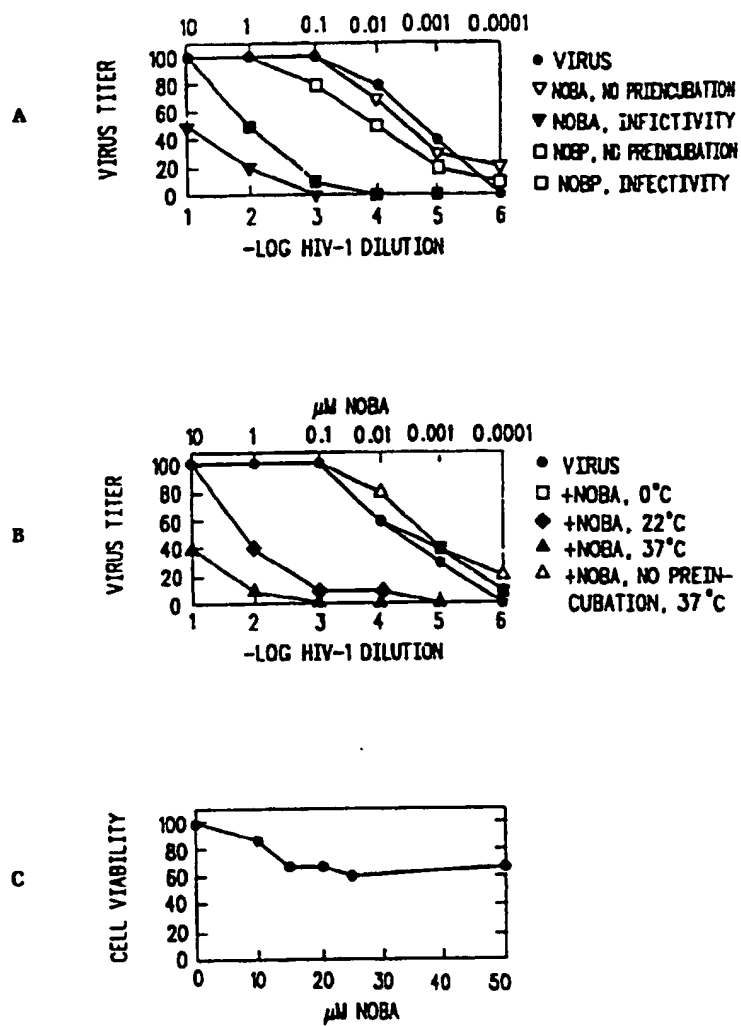


Figure 7A, 7B, 7C

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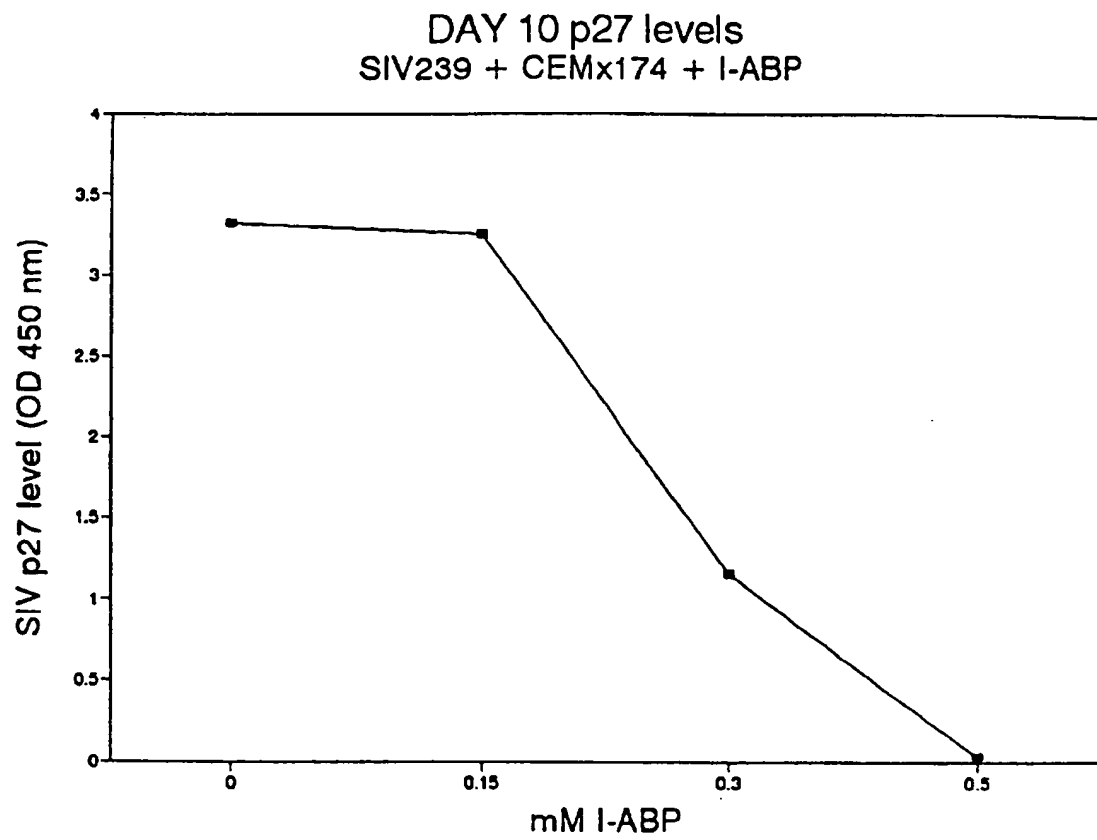


Figure 8

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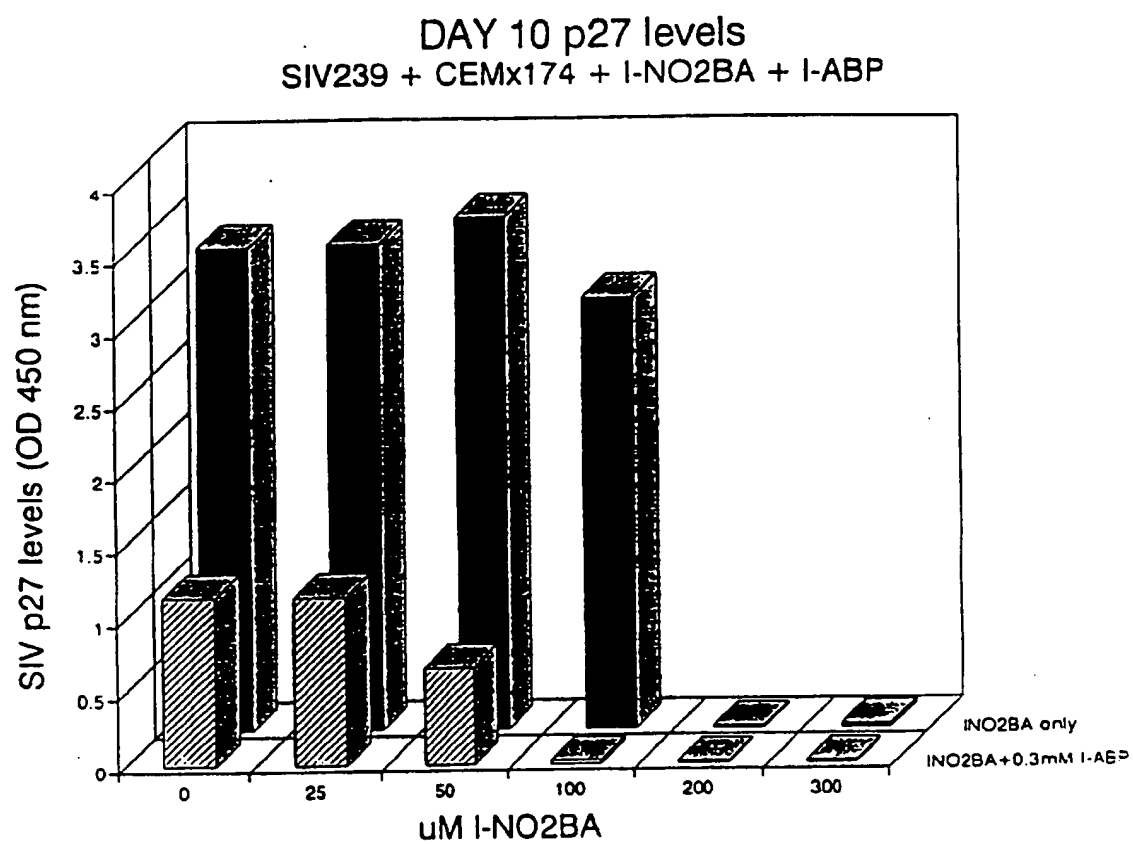


Figure 9A

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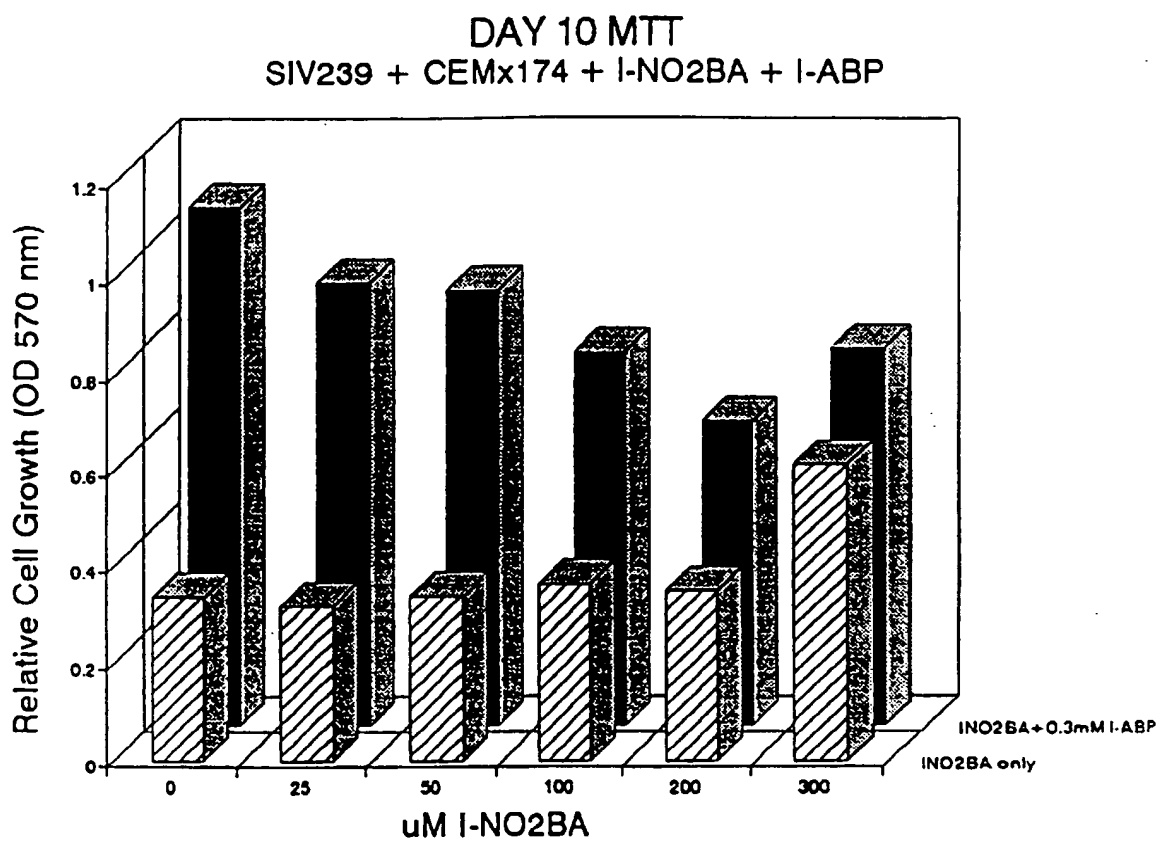


Figure 9B

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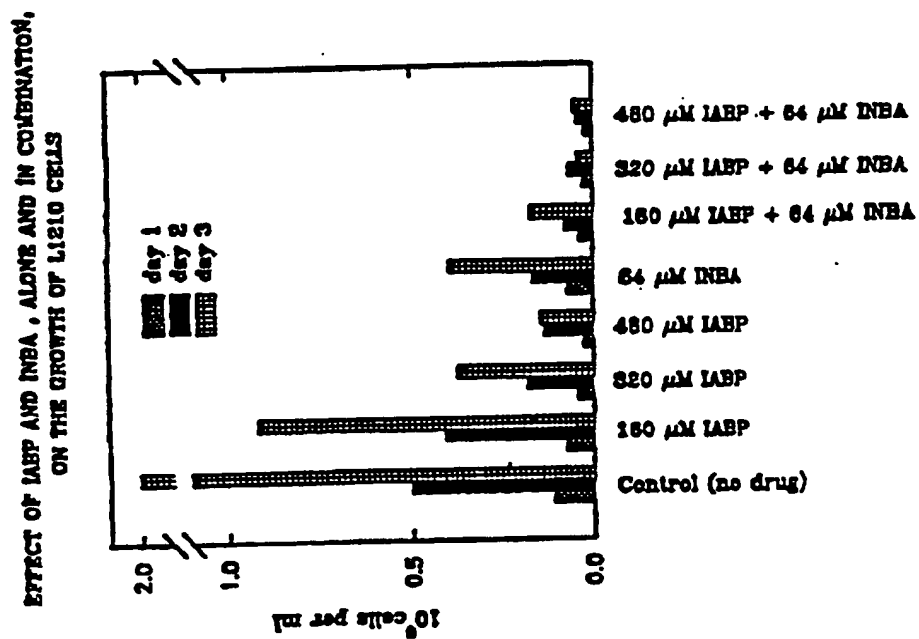


Figure 10

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 96/00420

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A61K45/06 A61K31/37 A61K31/225 A61K31/195 //(A61K31/37, 31:37, 31:225, 31:195), (A61K31/225, 31:165), (A61K31/195, 31:165)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	BIOCHEM. PHARMACOL., vol. 50, no. 5, 1995, pages 705-714, XP002006606 J. MENDELEYEV ET AL.: "Potential chemotherapeutic activity of 4-iodo-3-nitrobenzamide"	1-6,9, 13,14, 16-22, 26,28
P,Y	see the whole document	1-42
Y	--- PROC. WEST. PHARMACOL. SOC., vol. 37, 1994, pages 117-119, XP002006607 A. J. CHUANG ET AL.: "Comparison of the cytotoxic and antiretroviral effects of 3-nitrosobenzamide and 4-iodo-3-nitrobenzamide" see the whole document --- -/-	1-42

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
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Date of the actual completion of the international search

25 June 1996

Date of mailing of the international search report

05.07.96

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Stierman, B

INTERNATIONAL SEARCH REPORT

Intern. Patent Application No.

PCT/US 96/00420

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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Y	NATURE, vol. 361, no. 6411, 1993, pages 473-475, XP002006609 W.G. RICE ET AL.: "Inhibition of HIV-1 infectivity by zinc-ejecting aromatic C-nitroso compounds" cited in the application see the whole document ---	1-42
Y	PROC. NATL. ACAD. SCI., vol. 89, no. 16, 1992, pages 7703-7707, XP002006610 W.G. RICE ET AL.: "Induction of endonuclease-mediated apoptosis in tumor cells by C-nitroso-substituted ligands of poly(ADP-ribose) polymerase" cited in the application see the whole document ---	1-42
Y	FEBS LETT., vol. 326, no. 1-3, 1993, pages 140-144, XP002006611 A.J. CHUANG ET AL.: "Inhibition of the replication of native and 3'-azido-2',3'-dideoxy-thymidine(AZT)-resi- stant simian immunodeficiency virus (SIV) by 3-nitrosobenzamide" cited in the application see the whole document ---	1-42
Y	WO,A,94 26730 (OCTAMER INC) 24 November 1994 see the whole document ---	1-42
Y	WO,A,94 09776 (OCTAMER INC ;UNIV EMORY (US)) 11 May 1994 see the whole document ---	1-42
Y	WO,A,92 06687 (UNIV CALIFORNIA) 30 April 1992 see the whole document ---	1-42
1 Y	WO,A,92 18123 (OCTAMER INC) 29 October 1992 see the whole document ---	1-42
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INTERNATIONAL SEARCH REPORT

Internat'l Application No

PCT/US 96/00420

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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Y	W0,A,89 07441 (UNIV CALIFORNIA) 24 August 1989 see the whole document ---	1-15, 29-42
Y	BIOCHEM. BIOPHYS. RES. COMMUN., vol. 180, no. 2, 1991, pages 504-514, XP002006612 G.A. COLE ET AL.: "Inhibition of HIV-1 IIIB replication in AA-2 and MT-2 cells in culture by two ligands of poly (ADP-ribose) polymerase: 6-Amino-1,2-benzopyrone and 5-iodo-6-amino-1,2-benzopyrone" cited in the application see the whole document ---	1-15, 29-42
Y	EXP. CELL RES., vol. 193, no. 1, 1991, pages 1-4, XP002006613 E. KIRSTEN ET AL.: "Suppression of dexamethasone-stimulated DNA synthesis in an oncogene construct containing rat cell line by a DNA site-oriented ligand of poly-ADP-ribose polymerase: 6-Amino-1,2-benzopyrone" see the whole document -----	1-15, 29-42

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 96/00420

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WO-A-8907441	24-08-89	NONE	
